



Review

Comparative evaluation of bioactive compounds of various cultivars of pomegranate (*Punica granatum*) in different world regions

Shahindokht Bassiri-Jahromi^{1,*} and Aida Doostkam²

¹ Department of Medical Mycology, Pasteur Institute of Iran

² Department of pharmacology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran

* **Correspondence:** Email: bassiri@pasteur.ac.ir; Tel: +989122972237.

Abstract: Pomegranates (*Punica granatum*) are a significant source of bioactive compounds and have been widely used in traditional Eastern medicine in different countries and cultures. Pomegranate peel and pulp have significantly *antioxidant* capacity, and pomegranate pulp contain markedly anti-atherogenic activity against atherosclerosis diseases in humans and mice model investigations. Pomegranates from different cultivars possess different bioactive molecules, increasing the desired antioxidant properties genes and chemical profiles. This study provides an overview of geographic origins and an evaluation of the chemical analysis of flavonoids and polyphenolic content of the pomegranate from the local (Persian) and foreign pomegranate cultivars. The present narrative review aims to compare chemical analysis of total phenolic and flavonoids content of different pomegranate cultivars grown in various geographic regions in the world. This review demonstrated that remarkable differences existed in all factors rates measured in various cultivars, being considerably related to many factors such as cultivar type, growing region, maturity, cultivation, climate, edaphic condition, and storage situation. The data would be useful to produce better crops with high nutritional quality.

Keywords: pomegranate; polyphenols; flavonoids; phytomedicine

Abbreviations: GAE/g: Gallic acid equivalents per one gram; CE/g: Catechin equivalents (CE) per one gram; RE/g: Rutin equivalents per one gram; QuE: Quercetin equivalent

1. Introduction

Pomegranate is native to Persia and Mediterranean zone and has been widely used in many countries and cultures [1]. Pomegranate has attracted considerable attention for its health benefits in recent years. Results show that pomegranate juice has markedly high total phenolic contents and antioxidant capacity, being responsible for beneficial activities of pomegranates. Total phenolic content plays a probable role in preventing different diseases related to oxidative stress such as cardiovascular [2–4], cancer [5,6] and neurodegenerative diseases [7,8]. Pomegranate juice has excellent antioxidant activity and is beneficial for atherosclerosis prevention [2–4]. In this regard, polyphenols are capable of moderating the broad range of enzymes activities and cell receptors [9]. The main part of pomegranate seed oil is Punicic acid having anti-atherogenic effects [10].

Pomegranate extract has potential to decrease the incidence of collagen-induced arthritis. In an in-vivo study into mice fed by pomegranate extract, it was indicated that joint inflammation, arthritis severity and IL-6 level were decreased remarkably [11]. Pomegranate has been widely accepted for its antimicrobial [12–14], and anti-candidial activities by in-vitro [15] and iv-vivo investigations [16]. Crude extract of pomegranate peel yielded a compound that demonstrated potent antifungal activity to *Candida spp.* [13]. The compound was recognized as punicalagin, based on spectral analyses [17]. Punicalagin showed high activity against *Candida albicans* and *Candida parapsilosis*, as tested by minimum inhibitory concentration (MIC) [14]. Pomegranate pericarp and peel extracts are reported to possess strong activity against *Candida spp.*, with MICs of 125 µg/mL [18]. Salazar Aranda et al. [19] showed that polyphenolics compound had antioxidant activity, and they revealed its highest activity against *C. glabrata* by MIC test. The effect of their hydroxyl groups on their activity against *C. glabrata* is considerable. Furthermore, it has been shown that pomegranate peel has a high inhibition capacity against *C. albicans* [19,20]. Bassiri-Jahromi et al. in their in-vitro [15] and in-vivo [16] investigations indicated that pomegranate peel extract had potential antifungal activity against 5 various *Candida* species. This investigation demonstrated that among 8 different Persian pomegranate cultivars, Saveh sour malas peel extract indicated strongest antifungal activity against *C. albicans*, which was comparable to nystatin. Bassiri-Jahromi et al. reported that pomegranate peel extract had no adverse effects following application in the rats' model. Pomegranate peel extract application was effective and safe in treating oral candidiasis in the Wistar rats [21].

Schubert et al. (1999) in their studies clearly demonstrated that pomegranate fermented juice and seed procurement and pomegranate seed oil contained powerful antioxidant properties [22]. Therefore, pomegranate can play a potential role as natural food preservative, health promotion and therapeutic agents.

This paper provides a general review of the evaluation of polyphenolic and flavonoid contents of various pomegranate cultivars in different regions of the world.

2. Pomegranate polyphenols and flavonoid compound

Pomegranate has considerable content of phytochemicals compounds such as punicalagin, ellagitannins, anthocyanins, tannins, hydrolysable tannins, and punicic acid [18–23]. Pomegranate peel is a valuable source of polyphenolic compounds, known as punicalagin, which is an ellagitannin with antioxidant capacity and is unrivaled and unique to pomegranate [24].

Phenolic compounds have attracted increasing attention as agents for inhibiting and treating various oxidative stress correlated diseases, preventing conventional and novel biomarkers of tissue plasminogen activator (TPA) induced tumor promotion, as well as possessing chemo-preventive role in various tumor models [25]. These compounds are recognized for their attributes in scavenging free radicals and preventing in-vitro lipid oxidation [26,27].

Polyphenol is a significant antioxidant found in pomegranate seed and juice containing ellagitannin (punicalagin), gallic acid, ellagic acid, anthocyanins, catechins, caffeic acid, and quercetin [28]. Flavonoids may prevent coronary artery diseases by inducing various processes such as HDL increase, LDL decrease level, mast cell release reduction, and cardiovascular inflammation decrease. Flavonoids have been recognized with antiviral activity since the 1940s [29]. Selway et al. (1986) have reported that flavonoids contain antiviral activity against 11 types of viruses [30]. Furthermore, flavonoids possess protective effect against liver injury [31]. There is incisive documentary evidence that flavonoids have anti-mutagenic acting [32,33].

Pomegranate peel is a significant source of bioactive compounds such as flavonoids, polyphenols, ellagitannins, and proanthocyanidin [34]; however, this part of the fruit is inedible. The antibacterial activity of peel extracts of Indian Ganesh variety was tested by Malviya et al. using the agar well diffusion method against four bacterial strains, *Staphylococcus aureus*, *Salmonella typhi*, *Enterobacter aerogenes*, and *Klebsiella pneumoniae*. The pomegranate peel extracts showed significant antibacterial activities against all of the 4 bacterial strains tested [35].

The abundance of these compounds and their activities are related to cultivar type, climate, and growing region [36,37]. Up to now, polyphenols of different pomegranate cultivars in Iran [38], Turkey [39], the United States [40], Italy [41] and South Africa have been investigated [42]. Fawole et al. showed that the highest peel extract activity against monophenolase activity and phenolase activity was Bhagwa cultivar and Arakta cultivar with IC₅₀ values of 3.66 µg/mL and 15.88 µg/mL, respectively [43].

Almost 50% of the pomegranate weight corresponds to the peel [44]. Total polyphenols, flavonoids and pro-athocyanidins contents are superior in pomegranate peel extract than in pomegranate pulp extract owing to their powerful antioxidant capacity [45].

According to Shams Ardekani et al. (2011) [38] report from Iran, sour summer cultivar peel extract has the highest antioxidant activity with 118.074 mg or 274.132 µmol trolox/g. Sour summer cultivar is a strong source of natural antioxidants, phenolic and flavonoid content and the peel of Sweet Saveh malas, Sour summer and Black peel cultivars are suitable sources of phenolic and flavonoid compounds.

This review will investigate the evidence for the identity of the antioxidant content of various cultivars of pomegranate such as polyphenols and flavonoids, playing a probable role in preventing different diseases associated with oxidative stress such as cardiovascular, cancer and neurodegenerative diseases.

The results provide significant information about the compound of polyphenols and antioxidant content of different cultivars of pomegranate, which can be useful for expanding fruit processing professions and choosing favorable pomegranate genotypes to provide commercial agriculture.

The phenolic and flavonoids content are different; the antioxidant activity of various solvent extracts from pomegranate peel was also surveyed using in vitro assays.

Gil et al. (2000) [27] evaluated antioxidant acting of pomegranate by four comparative assays: ABTS, DPPH, DMPD, FRAP, and they were detected and quantified using ellagic acid anthocyanins,

and hydrolyzable tannins in pomegranate juice. They reported that the antioxidant capacity of commercial pomegranate juice was three times superior to red wine and green tea.

Pomegranate peels have significant superior antioxidant potency compared to other parts of pomegranate against free radical activities. It also contains higher total polyphenols, flavonoids and proanthocyanidins than pulp extract. Strong antioxidant potency of pomegranate peel extract may be due to its major polyphenolics contained [46]. In the present study, various cultivars from different regions were described evaluate the phenolic and flavonoid contents. Tables 1, 2 and 3 present the obtained data.

Table 1. Comparative evaluation of polyphenolic and flavonoid content of peels of various pomegranate (*Punica granatum*) cultivars from different regions in the world.

No.	Author/References	Cultivar	Region of growth (Country)	Fruit part	Total Polyphenolic Compound mg GAE/g extract	Flavonoids (mg/g), Total flavonoids mg CE/g extract
1	Fawole OA et al. (2012) [67]	Ganesh	South Africa	Peel	295.5 ± 23.91 d mg GAE/ g DM	121.1 ± 3.12 c mg CAE/g DM
2	Shiban MS et al. (2012) [68]	Yemeni variety	Yemen	Peel	274.1 ± 17.2 mg GAE/g	56.4 ± 2.7 c mg (RE)/g.
3	Ardekani MRS et al. (2011) [38]	Black peel	Iran	Peel	250.13 ± 33.03 mg GAE/g	36.40 ± 1.34 mg CAE/g DM
4	Kulkarni AP et al. (2004) [58]	Kashmir	India	Peel	249.4 mg GAE/g	59.4 mg CE/g dry solids
5	Li Y et al. (2006) [45]	China	China	Peel	249.4 ± 17.2 mg/g	59.1 ± 4.8 (mg/g)
6	Ardekani MRS et al. (2011) [38]	Sour Summer	Iran	Peel	226.56 ± 18.98 mg GAE/g	35.92 ± 0.84 mg CAE/g DM
7	Fawole OA et al. (2012) [67]	Bhagwa	South Africa	Peel	224.1 ± 6.86 c mg GAE/g DM	112.6 ± 1.51 b mg CAEg DM
8	Fawole OA et al. (2012) [67]	Ruby	South Africa	Peel	218.2 ± 4.53 bc mg GAE/g DM	126.0 ± 0.57 c mg CAE/g DM
9	Ardekani MRS et al. (2011) [38]	Saveh sour malas	Iran	Peel	216.74 ± 19.01 mg GAE/g	34.71 ± 1.34 mg CAE/g DM
10	Nasr CB et al. (1996) [69]	Tunesia	Tunesia	Peel	216.9 ± 7.3 mg GAE/g	
11	Mphahlelea RR et al. (2016) [66]	South Africa	South Africa	peel	215.21 ± 21.90 b	36.67 ± 3.43 ab
12	Mphahlelea RR et al. (2016) [66]	South Africa	South Africa	Peel & seed	138.36 ± 2.27 c	50.39 ± 6.93 a
13	Mphahlelea RR et al. (2016) [66]	South Africa	South Africa	Whole fruit	185.73 ± 3.89 b	23.35 ± 2.07 b
14	Fawole OA et al. (2012) [67]	Herskawitz	South Africa	Peel	198.1 ± 9.22 abc mg GAE/g DM	101.0 ± 1.02a mg CAE/g DM
15	Ardekani MRS et al. (2011) [38]	North white peel	Iran	Peel	192.72 ± 15.45 mg GAE/g	26.94 ± 0.48 mg CAE/g DM

Continued on next page

No.	Author/References	Cultivar	Region of growth (Country)	Fruit part	Total Polyphenolic Compound mg GAE/g extract	Flavonoids (mg/g), Total flavonoids mg CE/g extract
16	Fawole OA et al. (2012) [67]	Wonderful	South Africa	Peel	189.1 ± 3.79 ab mg GAE/g	97.8 ± 2.10a mg GAE/g DM
17	Fawole OA et al. (2012) [67]	Arakta	South Africa	Peel	187.4 ± 6.44ab mg GAE/g DM	103.0 ± 1.86a mg CAE/g DM
18	Rosas-Burgos EC (2017) [56]	Cultivar (PTO8)	Spain	Peel	187 ± 4 mg GAE/g	
19	Ardekani MRS et al. (2011) [38]	Sweet alac	Iran	Peel	184.10 ± 25.07 mg GAE/g	30.36 ± 2.44 mg CAE/g DM
20	Fawole OA et al. (2012) [67]	Molla de Elche	South Africa	Peel	179.3 ± 4.60a mg GAE/g DM	99.5 ± 2.94a mg CAE/g DM
21	Ardekani MRS et al. (2011) [38]	Agha mohammad ali	Iran	Peel	168.21 ± 13.9 mg GAE/g	33.52 ± 0.41 mg CAE/g DM
22	Belkacem N et al. (2014) [70]	Algerian pomegranate	Algeria	Peel	158.18 ± 0.66 mg GAE/g DM	12.8 ± 2.2 mg CE/g
23	Negi P et al. (2003) [71]	Pomegranate varieties from India	India	Peel	124.3 mg GAE/g	49.1 mg CE/g dry solids
24	Ardekani MRS et al. (2011) [38]	Sweet malas	Iran	Peel	121.11 ± 8.69 mg GAE/g	18.61 ± 0.53 mg CAE/g DM
25	Ardekani MRS et al. (2011) [38]	Sour white peel	Iran	Peel	98.24 ± 4.81 mg GAE/g	28.30 ± 0.54 mg CAE/g DM
26	Al-Rawahi AS et.al. (2014) [72]	Hellow	Oman	Peel	64.2 mg GAE/g	1.4 mg CE/g
27	Dipnaik HS et al. (2014) [73]	Indian pomegranate	India	Peel	59.73 ± 0.46 mg GAE/g	
28	Gozlekci S et al. (2011) [74]	Lefan	Turkish	Peel	3547.8a µg GAE/g fw	
31	Gozlekci S et al. (2011) [74]	Katirbasi	Turkish	Peel	3127.0b µg GAE/g	
34	Gozlekci S et al. (2011) [74]	Cekirdeksiz-IV	Turkish	Peel	2537.1c µg GAE/g fw. GAE/g fw	
35	Gozlekci S et al. (2011) [74]	Asinar	Turkish	Peel	17.75.4d GAE/g fw	
36	Souleman AMA et al. (2016) [59]	PG1	Egypt	Peel	172.4 ± 1.11c (GAE mg/g FW)	34.28 ± 1.47 (REmg/g FW)
37	Souleman AMA et al. (2016) [59]	PG2	Egypt	Peel	135.8 ± 0.92 (GAE mg/g FW)	29.65 ± 0.59a (REmg/g FW)
38	Souleman AMA et al. (2016) [59]	PG3	Egypt	Peel	98.6 ± 1.13a (GAE mg/g FW)	21.72 ± 0.38 (REmg/g FW)
39	Souleman AMA et al. (2016) [59]	PG4	Egypt	Peel	102.9 ± 1.28	30.29 ± 1.29a (REmg/g FW)
40	Souleman AMA et al. (2016) [59]	PG5	Egypt	Peel	95.8 ± 1.19a (GAE mg/g FW)	26.35 ± 1.16 (REmg/g FW)

Table 2. Comparative evaluation of polyphenolic and flavonoid content of juices of various pomegranate (*Punica granatum*) cultivars from different regions.

No.	Author/References	Cultivar	Region of growth (Country)	Total Polyphenolic Compound mg GAE/g extract	Flavonoids (mg/g), Total flavonoids mg CE/g extract
1	Gozlekci S et al. (2011) [74]	Lefan	Turkish	1551.5 µg GAE/g fw	
2	Gozlekci S et al. (2011) [74]	Katirbasi	Turkish	1229.5 mg/L µg GAE/g fw	
3	Gozlekci S et al. (2011) [74]	Asinar	Turkish	1307.3 µg GAE/g fw	
4	Nunzio MD et al. (2013) [75]	Hershkovitz	Italian	2057.2 ± 174.0a µg GAmL ⁻¹)	
5	Li X et al. (2015) [76]	XJ-TSL	China	4.352 ± 0.09 de d (GaE mg/mL)	0.118 ± 0.00 d QuE mg/mL
6	Li X et al. (2015) [76]	XJ-SSL	China	6.147 ± 0.11 b (GaE mg/mL)	0.045 ± 0.01 h QuE mg/mL
7	Li X et al. (2015) [76]	SD-TSL	China	7.429 ± 0.12 a (GaE mg/mL)	0.335 ± 0.13 a QuE mg/mL
8	Li X et al. (2015) [76]	SD-SSL	China	4.481 ± 0.11 d (GaE mg/mL)	0.093 ± 0.00 e QuE mg/mL
9	Li X et al. (2015) [76]	YN-SZ	China	3.234 ± 0.06 g (GaE mg/mL)	0.099 ± 0.00 e QuE mg/mL
10	Li X et al. (2015) [76]	YN-LZ	China	3.151 ± 0.05 g (GaE mg/mL)	0.084 ± 0.00 f QuE mg/mL
11	Li X et al. (2015) [76]	YN-SSL	China	4.142 ± 0.08 f (GaE mg/mL)	0.171 ± 0.00 c QuE mg/mL
12	Li X et al. (2015) [76]	SX-JPT	China	4.219 ± 0.10 ef (GaE mg/mL)	0.259 ± 0.00 b QuE mg/mL
13	Li X et al. (2015) [76]	SX-SBT	China	4.750 ± 0.08 c (GaE mg/mL)	0.170 ± 0.00 c QuE mg/mL
14	Li X et al. (2015) [76]	SX-SSL	China	4.735 ± 0.03 c (GaE mg/mL)	0.054 ± 0.00 g QuE mg/mL
15	Li X et al. (2015) [76]	SD-TSL	China	7.429 ± 0.12 a (GaE mg/mL)	0.335 ± 0.13 a QuE mg/mL
16	Souleman AMA et al. (2016) [59]	PG1	Egypt	72.4 ± 0.22 (GAE mg/g FW)	12.31 ± 0.91a (REmg/g FW)
17	Souleman AMA et al. (2016) [59]	PG2	Egypt	63.7 ± 1.16a (GAE mg/g FW)	9.64 ± 0.25 (REmg/g FW)
18	Souleman AMA et al. (2016) [59]	PG3	Egypt	64.3 ± 0.29a (GAE mg/g FW)	10.38 ± 1.34b (REmg/g FW)
19	Souleman AMA et al. (2016) [59]	PG4	Egypt	52.1 ± 0.18b (GAE mg/g FW)	10.68 ± 1.63b (REmg/g FW)

Continued on next page

No.	Author/References	Cultivar	Region of growth (Country)	Total Polyphenolic Compound mg GAE/g extract	Flavonoids (mg/g), Total flavonoids mg CE/g extract
20	Souleman AMA et al. (2016) [59]	PG5	Egypt	53.4 ± 0.32b (GAE mg/g FW)	12.91 ± 0.88a (REmg/g FW)
21	Zarei M et al. (2010) [63]	Aghaye	Iran	7.9749 ± 0.110c (mg/gr)	
22	Zarei M et al. (2010) [63]	Faroogh	Iran	7.2053 ± 0.150 c (mg/gr)	
23	Zarei M et al. (2010) [63]	Rabbab-e-Fars	Iran	7.8620 ± 0.200 c (mg/gr)	
24	Zarei M et al. (2010) [63]	Shahvar	Iran	5.2640 ± 0.120 c (mg/gr)	
25	Zarei M et al. (2010) [63]	Shirin-e-Bihaste)	Iran	5.6863 ± 0.110 c (mg/gr)	
26	Zarei M et al. (2010) [63]	Shirin-e-Mohali	Iran	5.6581 ± 0.110 c (mg/gr)	

Table 3. Comparative evaluation of polyphenolic and flavonoid content of seeds of various pomegranate (*Punica granatum*) cultivars from different regions.

No.	Author/References	Cultivar	Region of growth (Country)	Total Polyphenolic Compound mg GAE/g extract	Flavonoids (mg/g), Total flavonoids mg CE/g extract
1	Gozlekci S et al. (2011) [74]	Cekirdeksiz-IV	Turkish	117.0 µg GAE/g fw	
2	Gozlekci S et al. (2011) [74]	Katirbasi	Turkish	121.2 µg GAE/g fw	
3	Gozlekci S et al. (2011) [74]	Lefan	Turkish	125.3 µg GAE/g fw	
4	Souleman AMA et al. (2016) [59]	PG2	Egypt	95.6 ± 1.17a (GAE mg/g FW)	23.92 ± 1.34a (REmg/g FW)
5	Gozlekci S et al. (2011) [74]	Asinar	Turkish	177.4 µg GAE/g fw	
6	Souleman AMA et al. (2016) [59]	PG1	Egypt	123.7 ± 1.35 (GAE mg/g FW)	26.45 ± 0.29 (REmg/g FW)
7	Souleman AMA et al. (2016) [59]	PG3	Egypt	109.1 ± 0.68	22.59 ± 1.22 (REmg/g FW)
8	Souleman AMA et al. (2016) [59]	PG4	Egypt	92.8 ± 0.59a (GAE mg/g FW)	24.23 ± 0.95a (REmg/g FW)
9	Souleman AMA et al. (2016) [59]	PG5	Egypt	86.7 ± 1.26 (GAE mg/g FW)	19.84 ± 1.37 (REmg/g FW)

3. Discussion

Owing to the pomegranate health benefits, consumption of fresh pomegranates juice is increasing. Rich bioactive compound cultivars are a significant source of desirable antioxidant properties for health promotion. Pomegranate peel and pulp contain various kinds of antioxidants; however, pomegranate peel had the most antioxidant efficacy compared to the pulp and seed fractions [47].

The results of a number of investigations on phenolic compounds and antioxidant capacity of eighteen various pomegranate cultivars grown in Morocco revealed that the polyphenols concentration in pomegranate was high, and antioxidant activity and physico-chemical characteristics in pomegranates were influenced by the type of cultivar. Phenolic compounds of pomegranates are graded on phenolic acids (ellagic acid, gallic acid, chlorogenic acid, caffeic acid, vanillic acid, ferulic acids trans-2- Hydrocinnamic acid, quercetin). Additionally, some flavonoids such as catechin, rutin, quercetin and phloridzin were identified in pomegranate juice at various concentrations among the pomegranate cultivars [48].

Although pomegranate peels and the other remaining tissues are inedible, it would be possible to use them to prepare new products such as flavonoids capsules and other nutraceuticals after extraction. There are many pomegranate cultivars, which are classified and correlated based on some important parameters such as morphological characteristics of flower and tree.

Differences in the phenolic compound index among various parts of pomegranate were observed. Pomegranate peels indicated a high concentration of phenolic compounds, and ellagitannins have largest quantities in relation to pomegranate pulp and juice for each cultivar [49]. Owing to meeting the current market demand for fruits quality, the characteristics of pomegranate cultivars are important.

This paper provides an overview of biology evaluation of the phenolic and flavonoid contents of different pomegranate cultivars.

This review described a polyphenolic and flavonoid analysis and geographic origins of different pomegranate cultivars.

Different parts of the pomegranate such as peel can act as potentially antimicrobial agents. Table 1 shows that the Ganesh cultivar possesses the highest polyphenolic compound, indicating an association between polyphenols level and antibacterial activities. Bassiri-Jahromi et al. in their in-vivo investigation indicated that Saveh sour malas Persian cultivar possessed the best activity against 8 *Candida* strains [16]. Owing to the significant amount of phenolic compounds, Saveh sour malas is one of the best cultivars (Table 1). These findings demonstrated the relationship between the amount of polyphenolic compounds and its anti-candidiasis effect.

Therefore, it may be suggested as a natural alternative to synthetic antimicrobial agents. Punicalagin content in pomegranate extract is tannin, which is reported to be responsible for antimicrobial activity. Furthermore, the tannin rich bioactive fractions and ellagitannins have antibacterial [50], antifungal [51] and antimalaria properties [52].

Moreover, polyphenolic compounds not only play a role in controlling various related diseases to oxidative stress [9], but also regulate the activity of various cell receptors and enzymes [53].

Because the chemical composition of pomegranate peel is differ with the cultivar type such as sweet, sure, and sour-sweet [54,55], pomegranate antimicrobial activity may vary regarding its

cultivar [56]. Tehranifar et al. (2010) [57] reported that total polyphenols and tannins content in pomegranate juice were dependent on major chemical factors.

Kulkarni et al. [58] reported that antioxidant activity growth at the late-developmental phase was due to anthocyanins composition.

By investigating nine different cultivars, Shams Ardakani indicated that pulp of Sour Summer cultivar as a strong source of original antioxidants had the highest antioxidant activity than other cultivars ($p < 0.05$). The peel of Sour Summer, Sweet saveh malas, and Black peel is a considerable source of phenolic and flavonoid compound appropriate for phenolic and flavonoid purification and extraction. In addition, they reported that peel extracts had higher potential antioxidant activity and polyphenolic and flavonoid content than the pulps [38]. The pomegranate peel extract antioxidant capacity is 10 times greater than that of the pomegranate pulp extract. The North white peel and Black peel cultivar contain the highest flavonoid. Souleman et al. (2016) reported that seed of Egyptian pomegranate cultivar (PG1) contained the most total phenolic and flavonoids compounds (Table 3) [59].

Bassiri-Jahromi et al. (2015) [15] indicated that the peel extract of Saveh malas cultivar had the most effective element compared to other cultivars against *Candida spp* by MIC test [15]. Table 1 indicate that peel of Saveh malas pomegranate cultivar possesses a significant source of polyphenolic and flavonoid content compared to other cultivars. Difference in the pomegranate chemical compound is related to the cultivar, growing region, maturity, cultivation, climate, and storage situation [48]. This difference is also correlated with the latitude, altitude and longitude of growing regions [60]. The anthocyanin level of pomegranate juice variation was attributed to diversities of cultivars and growing region and various maturity levels of the pomegranate [40]. The pomegranate juice color is a significant index for juice quality; it is originally related to anthocyanin concentration.

Middha et al. [61] reported that higher total flavonoid concentration of pomegranate juice was almost correlated with sweet and sour cultivar and growing area. Although pomegranate anthocyanin pigments concentration decreased during 100 days, a considerable decrease in acidity was found as the significant chemical factor for increased incidence in over-ripe fruits.

This review clearly indicated that pomegranate peel extract possessed more natural antioxidants and activity as a health supplement than the pomegranate pulp extract. Derakhshani et al. in their study showed that the pomegranate peel extract contained high levels of antioxidant activity compared to seeds and juice in three different cultivars of various regions of Iran [62].

Table 1 shows the comparative evaluation of phenolic and flavonoids compounds attributes to various peels of pomegranate cultivars grown in the world.

These diversities may be owing to variety of cultivar, climate, edaphic condition, different maturity level, and particularly tannin specification method. Total tannin concentration pomegranate spectroscopic analyses by Khanavi et al. (2013) [1] in Iran revealed that Black peel cultivar had the most hyperoside content in its pulp and peel. Hyperoside is identified as the significant flavonoid with respect to antioxidant activity. The results indicated that commercial pomegranate juice had significant phenolic quantity and antioxidant capacity. Furthermore, Zarei et al. (2010) revealed a significant difference in total phenolics, tannins, and antioxidant activity in the juice of six different Persian cultivars (Table 2) [63]. The polyphenolic content and antioxidant potent of the whole pomegranate juice were remarkably superior to aril juice of pomegranate from the same cultivar, due to the entrance of phenolic compounds from the rind sections of pomegranate to the juice [64].

Hajimahmoodi et al. [65] reported that pulp of Sour summer cultivar had the most antioxidant potent among the nine various pomegranate cultivars. The antioxidant capacity of pomegranate peel extract was 10 times higher than the pulp extract.

The antioxidant activity of pomegranate peels showed a rapid decrease in 20 to 60 days of fruit development (13%) [58]. Furthermore, Kulkarni et al. [58] reported a slight but important decrease in anthocyanin pigment after 100 days (9.3%). Moreover, they concluded that the anthocyanin increasing and phenolics decreasing were correlated with each other. In this regard, phenolics were exhausted when the anthocyanin pigment formation and the phenols were destroyed and their contents reduced [56].

Mphahlele et al. in their investigation showed that freeze-drying could be a viable method to proceed pomegranate peel to maintain the maximum natural value of their bioactive compounds [66].

4. Conclusion

Different pomegranate cultivars had different polyphenol compositions and antioxidant potential. It is considerably associated with many factors such as cultivar type, growing region, maturity, cultivation, climate, edaphic condition, and storage situation [65]. They were also correlated with the latitude, altitude and longitude of growing regions. Additionally, difference in the average temperature and daily temperature during maturity and harvest period had significant effects on the total polyphenols, flavonoids and anthocyanin concentration of pomegranate.

Further future investigations are necessary to establish a database for pomegranate showing polyphenol and flavonoids compositions, antioxidant activities, physiochemical characteristics and their relation to environmental factors in various growing areas of the world. The data would be useful to produce better crops with higher nutritional quality. This database provides geographic product labeling and pomegranate brand identification.

Search strategy

This review is based on a method of systematic narrative review on comparative evaluation of bioactive compounds of various pomegranate cultivars. We conducted an extensive search using the PubMed, Web of Science, Science Direct, and Scopus databases in April 2018 to obtain related studies. This review aimed to present an overview of the comparison of the chemical analysis of total phenolic and flavonoids content of various pomegranate cultivars grown in different geographic regions of the world and to shows perspective advantages of pomegranate compound. It also aimed to summarize the present data from in vitro and in vivo tests animal trial systems and human clinical trials concerning the benefit of pomegranate compound. In addition, this review considered the pomegranate peel and pulp extracts activities and their future potential.

The findings of this review support that the pomegranate might be used in preventing and curing some diseases.

Research highlights

This study demonstrated considerable differences among the cultivars in most measured factors such as total poly phenolics, flavonoids, antioxidant activity and anthocyanins content. It also

highlights the current aspects and the new research into the potential therapeutic pomegranate for some diseases and the future of clinical research of pomegranate.

Acknowledgments

We gratefully acknowledge the financial support of the Pasteur Institute of Iran Research Council (project No.TP-9003).

Conflict of interest

No author of this paper has a conflict of interest, including specific financial interests, relationships, and/or affiliations relevant to the subject matter or materials included in this manuscript.

References

1. Khanavi M, Moghaddam G, Oveisi MR, et al. (2013) Hyperoside and anthocyanin content of ten different pomegranate cultivars. *Pak J Biol Sci* 16: 636–641.
2. Rosenblat M, Volkova N, Abassi Z, et al. (2015) High intrinsic aerobic capacity and pomegranate juice are protective against macrophage atherogenicity: Studies in high-vs. low-capacity runner (HCR vs. LCR) rats. *J Nutr Biochem* 26: 1015–1021.
3. Kaplan M, Hayek T, Raz A, et al. (2001) Pomegranate juice supplementation to atherosclerotic mice reduces macrophage lipid peroxidation, cellular cholesterol accumulation and development of atherosclerosis. *J Nutr* 131: 2082–2089.
4. Aviram M, Rosenblat M, Gaitini D, et al. (2004) Pomegranate juice consumption for 3 years by patients with carotid artery stenosis reduces common carotid intima-media thickness, blood pressure and LDL oxidation. *Clin Nutr* 23: 423–433.
5. Bassiri-Jahromi S (2018) *Punica granatum* (Pomegranate) activity in health promotion and cancer prevention. *Oncol Rev* 12: 345.
6. Middleton E, Kandaswami C, Theoharides TC (2000) The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. *Pharmacol Rev* 52: 673–751.
7. Rojanathammanee L, Puig KL, Combs CK (2013) Pomegranate polyphenols and extract inhibit nuclear factor of activated T-cell activity and microglial activation in vitro and in a transgenic mouse model of Alzheimer disease. *J Nutr* 143: 597–605.
8. Tapias V, Cannon JR, Greenamyre JT (2014) Pomegranate juice exacerbates oxidative stress and nigrostriatal degeneration in Parkinson's disease. *Neurobiol Aging* 35: 1162–1176.
9. Scalbert A, Manach C, Morand C, et al. (2005) Dietary polyphenols and the prevention of diseases. *Crit Rev Food Sci Nutr* 45: 287–306.
10. Zarfeshany A, Asgary S, Javanmard SH (2014) Potent health effects of pomegranate. *Adv Biomed Res* 3: 100.
11. Shukla M, Gupta K, Rasheed Z, et al. (2008) Consumption of hydrolyzable tannins-rich pomegranate extract suppresses inflammation and joint damage in rheumatoid arthritis. *Nutrition* 24: 733–743.

12. Betanzos-Cabrera G, Montes-Rubio PY, Fabela-Illescas HE, et al. (2015) Antibacterial activity of fresh pomegranate juice against clinical strains of *Staphylococcus epidermidis*. *Food Nutr Res* 59: 27620.
13. Ferrazzano GF, Scioscia E, Sateriale D, et al. (2017) In vitro antibacterial activity of Pomegranate juice and peel extracts on cariogenic bacteria. *Biomed Res Int* 2017: 2152749.
14. Duman AD, Ozgen M, Dayisoğlu KS, et al. (2009) Antimicrobial activity of six pomegranate (*Punica granatum* L.) varieties and their relation to some of their pomological and phytonutrient characteristics. *Molecules* 14: 1808–1818.
15. Bassiri-Jahromi S, Katiraei F, Hajimahmoodi M, et al. (2015) In vitro antifungal activity of various Persian cultivars of *Punica granatum* L. extracts against *Candida* species. *Jundishapur J Nat Pharm Prod* 10: e19754.
16. Bassiri-Jahromi S, Pourshafie MR, Mirabzadeh-Ardakani E, et al. (2018) In vivo comparative evaluation of pomegranate (*Punica granatum*) peel extract as alternative agents for nystatin against oral candidiasis. *Iran J Med Sci* 43: 296–304.
17. Endo EH, Cortez DA, Ueda-Nakamura T, et al. (2010) Potent antifungal activity of extracts and pure compound isolated from pomegranate peels and synergism with fluconazole against *Candida albicans*. *Res Microbiol* 161: 534–540.
18. Anibal PC, Peixoto ITA, Foglio MA, et al. (2013) Antifungal activity of the ethanolic extracts of *Punica granatum* L. and evaluation of the morphological and structural modifications of its compounds upon the cells of *Candida* spp. *Braz J Microbiol* 44: 839–848.
19. Salazar-Aranda R, Granados-Guzman G, Perez-Meseguer J, et al. (2015) Activity of Polyphenolic Compounds against *Candida glabrata*. *Molecules* 20: 17903–17912.
20. Madugula P, Reddy S, Koneru J, et al. (2017) “Rhetoric to Reality”—Efficacy of *Punica Granatum* Peel Extract on Oral Candidiasis: An in vitro Study. *J Clin Diagn Res* 11: ZC114–ZC117.
21. Bassiri-Jahromi S, Pourshafie MR, Mirabzadeh E, et al. (2015) *Punica granatum* Peel Extract Toxicity in Mice. *Jundishapur J Nat Pharm Prod* 10: e23770.
22. Schubert SY, Lansky EP, Neeman I (1999) Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *J Ethnopharmacol* 66: 11–17.
23. Mahboubi A, Asgarpanah J, Sadaghiyani PN, et al. (2015) Total phenolic and flavonoid content and antibacterial activity of *Punica granatum* L. var. *pleniflora* flowers (Golnar) against bacterial strains causing foodborne diseases. *BMC Complement Altern Med* 15: 366.
24. Turrini E, Ferruzzi L, Fimognari C (2015) Potential effects of pomegranate polyphenols in cancer prevention and therapy. *Oxid Med Cell Longevity* 2015: 938475.
25. Afaq F, Saleem M, Krueger CG, et al. (2005) Anthocyanin- and hydrolyzable tannin-rich pomegranate fruit extract modulates MAPK and NF-kappa B pathways and inhibits skin tumorigenesis in CD-1 mice. *Int J Cancer* 113: 423–433.
26. Li AN, Li S, Zhang YJ, et al. (2014) Resources and Biological Activities of Natural Polyphenols. *Nutrients* 6: 6020–6047.
27. Gil MI, Tomas-Barberan FA, Hess-Pierce B, et al. (2000) Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *J Agric Food Chem* 48: 4581–4589.
28. Noda Y, Kaneyuki T, Mori A, et al. (2002) Antioxidant activities of pomegranate fruit extract and its anthocyanidins: Delphinidin, cyanidin, and pelargonidin. *J Agric Food Chem* 50: 166–171.

29. Viuda-Martos M, J. Fernandez-Lopez J, J.A. Perez-Alvarez JA (2010) Pomegranate and its many functional components as related to human health: A review. *Compr Rev Food Sci Food Saf* 10: 635–654.
30. Selway JW, (1986) Antiviral activity of flavones and flavans, In: Cody V, Middleton E, Harborne JB (Eds.), *Plant Flavonoids in Biology and Medicine: Biochemical, Pharmacological, and Structure-Activity Relationships*, Alan R. Liss Inc., New York, 521–536.
31. Perrissoud D, Weibel I (1980) Protective effect of (+)cyanidanol-3 in acute liver injury induced by galactosamine or carbon tetrachloride in the rat. *Naunyn Schmiedebergs Arch Pharmacol* 312: 285–291.
32. Cano-Lamadrid M, Marhuenda-Egea FC, Hernandez F, et al. (2016) Biological Activity of Conventional and Organic Pomegranate Juices: Antioxidant and Antimutagenic Potential. *Plant Foods Hum Nutr* 71: 375–380.
33. Elwej A, Ben Salah G, Kallel C, et al. (2016) Protective effects of pomegranate peel against hematotoxicity, chromosomal aberrations, and genotoxicity induced by barium chloride in adult rats. *Pharm Biol* 54: 964–974.
34. Sreekumar S, Sithul H, Muraleedharan P, et al. (2014) Pomegranate fruit as a rich source of biologically active compounds. *Biomed Res Int* 2014: 686921.
35. Malviya S, Arvind, Jha A, et al. (2014) Antioxidant and antibacterial potential of pomegranate peel extracts. *J Food Sci Technol* 51: 4132–4137.
36. Melgarejo P, Salazar DM, Artes F (2000) Organic acids and sugars composition of harvested pomegranate fruits. *Eur Food Res Technol* 211: 185–190.
37. Poyrazoglu E, Gokmen V, Artik N (2002) Organic acids and phenolic compounds in pomegranates (*Punica granatum* L.) grown in Turkey. *J Food Compos Anal* 15: 567–575.
38. Ardekani MRS, Hajimahmoodi M, Oveisi MR, et al. (2011) Comparative Antioxidant Activity and Total Flavonoid Content of Persian Pomegranate (*Punica granatum* L.) Cultivars. *Iran J Pharm Res* 10: 519–524.
39. Ozcelik B, Balaban N, Aksaray S, et al. (2006) In vitro susceptibility of *Candida* spp. Isolated from clinical specimens against some antifungal agents. *Turkish J Pharm Sci* 3: 1–6.
40. Young JE, Pan Z, Teh HE, et al. (2017) Phenolic composition of pomegranate peel extracts using a liquid chromatography-mass spectrometry approach with silica hydride columns. *J Sep Sci* 40: 1449–1456.
41. Fanali C, Belluomo MG, Cirilli M, et al. (2016) Antioxidant activity evaluation and HPLC-photodiode array/MS polyphenols analysis of pomegranate juice from selected italian cultivars: A comparative study. *Electrophoresis* 37: 1947–1955.
42. Fawole OA, Opara UL (2013) Harvest discrimination of pomegranate fruit: postharvest quality changes and relationships between instrumental and sensory attributes during shelf life. *J Food Sci* 78: S1264–S1272.
43. Fawole OA, Makunga NP, Opara UL (2012) Antibacterial, antioxidant and tyrosinase-inhibition activities of pomegranate fruit peel methanolic extract. *BMC Complement Altern Med* 12: 200.
44. Sreekumar S, Sithul H, Muraleedharan P, et al. (2014) Pomegranate Fruit as a Rich Source of Biologically Active Compounds. *Biomed Res Int* 2014: 686921.
45. Li Y, Guo C, Yang J, et al. (2006) Evaluation of antioxidant properties of pomegranate peel extract in comparison with pomegranate pulp extract. *Food Chem* 96: 254–260.

46. Apak R, Guclu K, Demirata B, et al. (2007) Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. *Molecules* 12: 1496–1547.
47. Orak HH, Yagar H, Isbilir SS (2012) Comparison of antioxidant activities of juice, peel, and seed of pomegranate (*Punica granatum* L.) and inter-relationships with total phenolic, Tannin, anthocyanin, and flavonoid contents. *Food Sci Biotechnol* 21: 373–387.
48. Hmid I, Elothmani D, Hanine H, et al. (2017) Comparative study of phenolic compounds and their antioxidant attributes of eighteen pomegranate (*Punica granatum* L.) cultivars grown in Morocco. *Arabian J Chem* 10: S2675–S2684.
49. Russo M, Fanali C, Tripodo G, et al. (2018) Analysis of phenolic compounds in different parts of pomegranate (*Punica granatum*) fruit by HPLC-PDA-ESI/MS and evaluation of their antioxidant activity: Application to different Italian varieties. *Anal Bioanal Chem* 410: 3507–3520.
50. Reddy MK, Gupta SK, Jacob MR, et al. (2007) Antioxidant, antimalarial and antimicrobial activities of tannin-rich fractions, ellagitannins and phenolic acids from *Punica granatum* L. *Planta Med* 73: 461–467.
51. Foss SR, Nakamura CV, Ueda-Nakamura T, et al. (2014) Antifungal activity of pomegranate peel extract and isolated compound punicalagin against dermatophytes. *Ann Clin Microbiol Antimicrob* 13: 32.
52. Dell'Agli M, Galli GV, Bulgari M, et al. (2010) Ellagitannins of the fruit rind of pomegranate (*Punica granatum*) antagonize in vitro the host inflammatory response mechanisms involved in the onset of malaria. *Malar J* 9: 208.
53. Li AN, Li S, Zhang YJ, et al. (2014) Resources and biological activities of natural polyphenols. *Nutrients* 6: 6020–6047.
54. Labbe M, Ulloa PA, Lopez F, et al. (2016) Characterization of chemical compositions and bioactive compounds in juices from pomegranates ('Wonderful', 'Chaca' and 'Codpa') at different maturity stages. *Chilean J Agric Res*, 76.
55. Cam M, Hisil Y, Durmaz G (2009) Characterisation of Pomegranate Juice From Ten Cultivars Grown in Turkey. *Int J Food Prop* 12: 388–395.
56. Rosas-Burgos EC, Burgos-Hernandez A, Noguera-Artiaga L, et al. (2017) Antimicrobial activity of pomegranate peel extracts as affected by cultivar. *Sci Food Agric* 97: 802–810.
57. Tehranifar A, Zarei M, Nematia Z, et al. (2010) Investigation of physico-chemical properties and antioxidant activity of twenty Iranian pomegranate (*Punica granatum* L.) cultivars. *Sci Hort* 126: 180–185.
58. Kulkarni AP, Aradhya SM (2005) Chemical changes and antioxidant activity in pomegranate arils during fruit development. *Food Chem* 93: 319–324.
59. Souleman AMA, Ibrahim GE (2016) Evaluation of Egyptian pomegranate cultivars for antioxidant activity, phenolic and flavonoid contents. *Egypt Pharm J* 15: 143–149.
60. Sheidai M, Kolahizadeh K, Noormohammadi Z, et al. (2012) Correlation between geography and cytogenetic diversity in Pomegranate (*Punica granatum* L.) cultivars. *Acta Botanica Brasiliica* 26: 948–957.
61. Middha SK, Usha T, Pande V (2013) HPLC Evaluation of phenolic profile, nutritive content, and antioxidant capacity of extracts obtained from *punica granatum* fruit peel. *Adv Pharmacol Sci* 3: 296236.

62. Derakhshani Z, Ferrante M, Tadi M, et al. (2018) Antioxidant activity and total phenolic content of ethanolic extract of pomegranate peels, juice and seeds. *Food Chem Toxicol* 114: 108–111.
63. Zarei M, Azizi M, Bashiri-Sadr Z (2010) Studies on Physico-Chemical Properties and Bioactive Compounds of Six Pomegranate Cultivars Grown in Iran. *J Food Technol* 8: 112–117.
64. Akhavan HR, Barzegar M, Weidlich H, et al. (2015) Phenolic compounds and antioxidant activity of juices from ten iranian pomegranate cultivars depend on extraction. *J Chem* 2015: 7.
65. Hajimahmoodi M, Oveisi MR, Sadeghi N, et al. (2008) Antioxidant properties of peel and pulp hydro extract in ten Persian pomegranate cultivars. *Pak J Biol Sci* 11: 1600–1604.
66. Mphahlele RR, Fawole OA, Makunga NP, et al. (2016) Effect of drying on the bioactive compounds, antioxidant, antibacterial and antityrosinase activities of pomegranate peel. *BMC Complement Altern Med* 16: 143.
67. Fawole OA, Makunga NP, Opara UL (2012) Antibacterial, antioxidant and tyrosinase-inhibition activities of pomegranate fruit peel methanolic extract. *BMC Complement Altern Med* 12: 200.
68. Shibani MS, Al-Otaibi MM, Al-Zoreky NS (2012) Antioxidant activity of pomegranate (*Punica granatum* L.) fruit peels. *Food Nutr Sci* 3: 991–996.
69. Nasr CB, Ayed N, Metche M (1996) Quantitative determination of the polyphenolic content of pomegranate peel. *Z Lebensm Unters Forsch* 203: 374–378.
70. Belkacem N, Djaziri R, Lahfa F, et al. (2014) Phytochemical Screening and In Vitro Antioxidant Activity of Various *Punica granatum* L. Peel Extracts from Algeria: A comparative study. *Phytotherapie* 12: 372–379.
71. Negi P, Jayaprakasha G, Jena B (2003) Antioxidant and antimutagenic activities of pomegranate peel extracts. *Food Chem* 80: 393–397.
72. Al-Rawahi AS, Edwards G, Al-Sibani M, et al. (2014) Phenolic constituents of pomegranate peels (*Punica granatum* L.) cultivated in Oman. *Eur J Med Plants* 4: 315–331.
73. Dipnaik HS, Shrivastava P, Jacob SM (2014) *Aspergillus Niger* mediated Solid State Fermentation (SSF) of Pomegranate Peels, yields Bioactive Products. *World J Pharm Res* 3045: 1377–1394.
74. Gozlekci S, Saracoglu O, Onursal E, et al. (2011) Total phenolic distribution of juice, peel, and seed extracts of four pomegranate cultivars. *Pharmacogn Mag* 7: 161–164.
75. Nunzio MD, Toselli M, Verardo V, et al. (2013) Counteraction of oxidative damage by pomegranate juice: Influence of the cultivar. *J Sci Food Agric* 93: 3565–3573.
76. Li X, Wasila H, Liu L, et al. (2015) Physicochemical characteristics, polyphenol compositions and antioxidant potential of pomegranate juices from 10 Chinese cultivars and the environmental factors analysis. *Food Chem* 175: 575–584.



AIMS Press

© 2019 the Author(s), licensee AIMS Press. This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>)