



Review

Cell ageing: a flourishing field for neurodegenerative diseases

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Abstract: Cellular senescence is viewed as an irreversible cell-cycle arrest mechanism involving a complexity of biological progressive processes and the acquisition of diverse cellular phenotypes. Several cell-intrinsic and extrinsic causes (stresses) may lead to diverse cellular signaling cascades that include oxidative stress, mitochondrial dysfunction, DNA damage, excessive accumulation of misfolded proteins, impaired microRNA processing and inflammation. Here we review recent advances in the causes and consequences of brain cell ageing, including the senescence of endothelial cells at the central nervous system barriers, as well as of neurons and glial cells. We address what makes ageing an important risk factor for neurodegenerative disorders, such as Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis and cerebrovascular disease. In particular, we highlight the importance of defects in mitochondrial dynamics, in the cathepsin activity imbalance, in cell-cell communication, in the accumulation of misfolded and unfolded proteins and in the microRNA profiling as having potential impact on cellular ageing processes. Another important aspect is that the absence of specific senescence biomarkers has hampered the characterization of senescent cells in ageing and age-associated diseases. In accordance, the senescence-associated secretory phenotype (SASP) or secretome was shown to vary in distinct cell types and upon different stressors, and SASP heterogeneity is believed to create subsets of senescent cells. In addition to secreted proteins, we then place extracellular vesicles (exosomes and ectosomes) as important mediators of intercellular communication with pathophysiological roles in disease spreading, and as emerging targets for therapeutic intervention. We also discuss the application of engineered extracellular vesicles as vehicles for drug delivery. Finally, we summarize current knowledge on methods to rejuvenate senescent cells and we review cell replacement therapeutic strategies. A deeper understanding of the molecular mechanisms underlying the senescence of cells may open novel therapeutic approaches for age-related pathologies and for

extending healthy human life span.

Keywords: glia senescence; microRNA profiling; extracellular vesicles; senescence of endothelial cells; neuronal senescence; hallmarks of ageing; cathepsins; CX3CL1/CX3CR1 signaling; therapeutic opportunities

1. Introduction

Ageing is associated with a declined ability to handle cellular stress, including impaired mitochondrial function, overproduction of reactive oxygen species (ROS), DNA damage, excessive accumulation of misfolded proteins and impaired microRNA (miRNA) processing that renders the cell more susceptible to stress [1]. Emde and Hornstein [1] support that the biosynthesis of miRNAs, which are small non-coding RNAs that mediate post-transcriptional activity, are intimately linked with the cellular stress signaling pathways. Actually, miRNAs are considered to have great relevance in ageing as in many other cellular processes [2]. Cellular senescence by preventing unlimited cell proliferation may also have implications in tumor suppression [3]. When gross chromosomal alterations occur during mitosis, the generation of signals may either lead to apoptosis and necrosis or to cell senescence generally known as mitotic catastrophe [4]. This cellular senescence due to an irreversible growth arrest, known as replicative senescence, may then be triggered by various events related with inflammatory stress, progressive telomere erosion and decreased regeneration ability [5]. However, it is also suggested that cellular senescence may promote tissue repair and cancer progression [6]. Indeed, the senescent cells develop a senescent-associated secretory phenotype (SASP) whose factors (e.g. growth factors, proteases chemokines, interleukins, and extracellular matrix components) may favor the appearance of aggressive cancer cells phenotypes in the neighborhood or attract immune cells, as well as mobilize stem and progenitor cells for resolving tissue damage [6,7,8]. Nevertheless, cellular senescence and ageing are consistently considered risk factors for the pathogenesis of most neurodegenerative disorders, including Alzheimer's disease (AD) [9,10], Parkinson's disease (PD) [11,12], amyotrophic lateral sclerosis (ALS) [13,14], cerebrovascular disorders [15,16] and glaucoma [17]. For instance, neurovascular changes (comprising brain endothelial cells, pericytes, glia and neurons) and dysfunction of the blood-brain barrier (BBB) by ageing has been considered to eventually precede neuronal changes and the occurrence of neurodegenerative diseases, such AD [18]. Similarly, dysfunction of the blood-cerebrospinal fluid barrier (BCSFB) was suggested to relate with age-associated cognitive decline and neurodegenerative disease [19]. Ageing also contributes to an increased production of amyloid-beta (A β) peptide associated to AD [20], even considering that the amyloid cascade hypothesis is a controversial issue [21].

It is now accepted that the senescent cells are functionally altered and their number increase with ageing [22,23,24]. Cellular senescence occurs in culture and *in vivo* as a response to excessive extracellular or intracellular stress and was shown to accumulate along life, mainly in renewable tissues and during persistent inflammation [8]. Hallmarks of the senescent cells are increased expression of senescence-associated beta-galactosidase (SA- β -gal) [25] and p16^{INK4a}, commonly expressed by quiescent or terminally differentiated cells [6], together with the appearance of SASP and its associated robust secretion of factors [8].

Senescent cells increasingly release vesicles that have been commonly designed as exosomes and microvesicles or ectosomes (Figure 1). While exosomes may either activate or inhibit the immune system depending on the expressed proteins, ectosomes are suggested to have anti-inflammatory/immunosuppressive properties [26]. The packaging of miRNAs into vesicles are an important step to their delivery into target cells by plasma membrane fusion and endocytosis. Once they may be differently represented when released by senescent cells it is conceivable that such miRNAs can be implicated as detrimental factors in ageing [27].

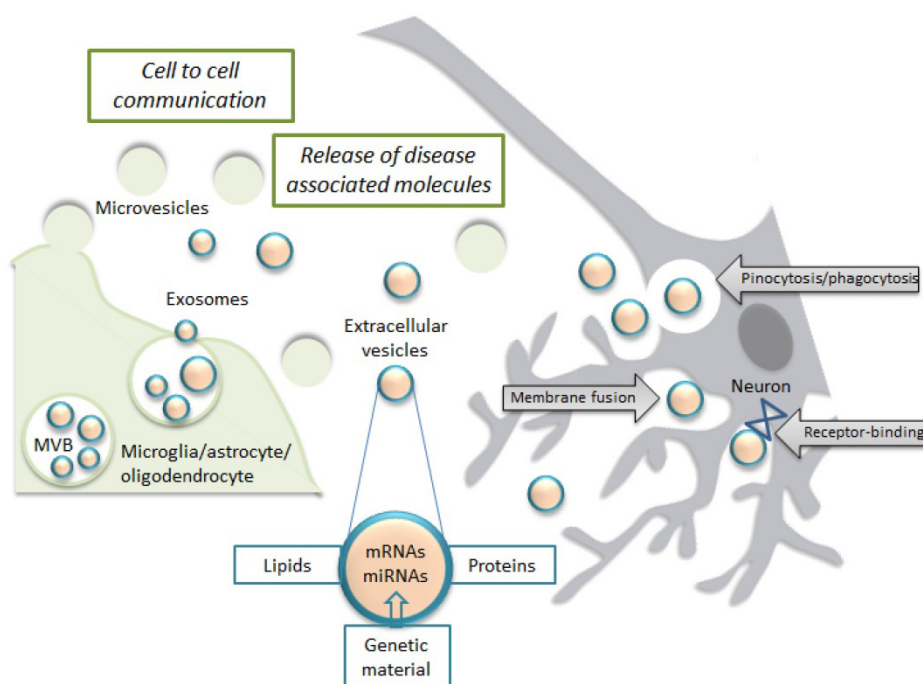


Figure 1. Biogenesis of extracellular vesicles, constituents and emerging roles in neuron-glia communication, cell senescence and disease spread. Ectosomes are produced directly from the plasma membrane, and exosomes from multivesicular bodies (MVB). Neuronal toxins or disease-associated molecules are processed by the cell and released to the extracellular space through ectosomes and exosomes.

Here we will summarize the common denominators of ageing (Table 1), the current knowledge on the cellular processes that predispose to neurodegeneration and we discuss the pathogenesis of age-related neurodegenerative diseases, including AD, PD, ALS and cerebrovascular disease. Linkage between cell senescence and deregulated signaling networks will be uncovered. We also give an overview on the existing knowledge on microvesicular secretion of miRNAs, their potential impact on cellular senescence, and implication in brain ageing and associated vulnerabilities to oxidative stress and inflammation. Finally, we will review recent information on interventions directed to rejuvenate senescent cells and on cell replenishment approaches.

Table 1. Candidate hallmarks and contribution to ageing.

Hallmarks of ageing	Causes	Relevance
Genomic instability	Increased DNA damage accumulation throughout life (delayed nuclear DNA repair, mutations and deletions in aged mitochondrial DNA, and defects in the nuclear architecture)	Relevance to normal ageing remains unsolved
Telomere attrition	Progressive and cumulative loss of telomere-protective sequences from chromosome ends, short telomere length and activation of DNA damage response (DDR)	Pathological telomere dysfunction accelerates ageing in rodents and humans
Epigenetic alterations	Alterations in DNA methylation patterns, post-translational modification of histones, chromatin remodeling and transcriptional alterations (non-coding RNAs, including a class of miRNAs designated by gero-miRs)	Understanding and modulating the epigenome may prevent/delay age-related disorders and extend healthy lifespan
Loss of proteostasis and impaired protein homeostasis	Chronic expression of unfolded, misfolded or aggregated proteins due to failure in restoring the structure of misfolded proteins and inability to remove/degrade such proteins	Accumulation of damaged components
Deregulated nutrient-sensing	Decline of the growth hormone and insulin-like growth factor, increase of mTOR, down regulation of AMPK and increase/decrease of sirtuins	Increased lifespan or healthspan by dietary restriction
Mitochondrial dysfunction	Increased production of reactive oxygen species, defective bioenergetics, alterations in mitochondrial dynamics by fission and fusion imbalance of events, changes in the lipid composition of mitochondrial membranes and defective quality control by mitophagy	Although severe mitochondrial dysfunction has a high impact in the processes of ageing, mild respiratory deficiencies may extend lifespan, possibly by hermetic response
Accumulation of senescent cells	Increased generation of senescent cells or decreased clearance rate of those cells, resulting from reduced immune response, or from acquisition of a senescence and proinflammatory associated secretory cell phenotype	Though cell senescence prevents the spread of damaged cells, their accumulation, by inefficient cell replacement mechanisms, may exacerbate damage and favor ageing
Stem cell exhaustion	Decline in the regenerative potential by deficient proliferation of stem and progenitor cells, or exhaustion of stem cell niches by excessive proliferation of those cells	Prevention of stem cell decline and cell rejuvenation to reverse the aging phenotype may halt exhaustion
Altered intercellular communication	Deregulated neurohormonal signaling, increased inflammatory signaling pathways and deteriorated cell-cell cross-talk	Restoration of defective cell-cell communication

Based on the works by López-Otín et al. [36], Morrison and Hof [37], Ren et al. [38] and Freund et al. [39].

2. Brain cell senescence: causes and consequences

Cellular senescence is characterized by an irreversible growth arrest and a combination of changes in cell morphology, function and behavior. It was first described as resultant of replicative senescence (RS), i.e. most cells cannot divide indefinitely as a consequence of finite number of cell doublings [28], even in the presence of nutrients and growth factors, but remain viable for weeks. Senescent cells are also induced by continued stress and damage from various stimuli, including DNA-damaging agents, oxidative stress, chemotherapeutic compounds and overexpression of oncogenes [29]. The senescent phenotype is characterized by irreversible growth arrest, apoptosis resistance and gene expression modification [30]. One of the first markers to be used in the identification of senescent cells was the SA- β -gal [25] due to increased lysosomal biogenesis in senescent cells [31,32]. Promising additional markers are the proteins acting as negative regulators of the cell cycle such as p15^{INK4b}, p16^{INK4a} and decoy death receptor-2 (DCR2) [33,34]. Others include the up-regulation of cathepsin D and down-regulation of eukaryotic translation elongation factor 1beta2 [35]. However, the search of definitive senescence biomarkers is still ongoing.

The first step toward senescence seems to involve the combined activity of two tumor tumor-suppressor proteins, p53 and retinoblastoma protein (pRB). Accumulating evidence indicates that the p53-p21 pathway is implicated in senescence due to telomere shortening or DNA damage, while p16-pRb pathway is partly involved in stress-induced premature senescence (SIPS) [29]. Both RS and SIPS are believed to create a barrier to tumorigenesis.

Cells undergoing senescence produce increased release of inflammatory cytokines [38] and SASP cells secrete interleukin(IL)-6, IL-8, IL-1 α , IL-1 β and metalloproteinases [6,8]. In addition, chronic inflammation creates a feedback loop in the immune system that leads to overproduction of cytokines and increased risk of deleterious effects [39]. Senescent cells accumulate with age and are present in sites of age-associated pathology, while contributing to age-related diseases [27,30]. Interestingly, the increase in p16 expression is associated with a reduced cell progenitor function and neurogenesis, thus contributing to reduced regenerative capacity in mammalian ageing [40].

2.1. Senescence of endothelial cells at CNS barriers

Senescence of endothelial cells was shown to compromise the integrity of barriers at the interfaces between the central nervous system (CNS) and the circulating blood, mainly by affecting tight and adherens junctions [41,42].

Although BBB is the most mentioned one, it is now recognized that BCSFB also has an important role by producing most of the cerebrospinal fluid (CSF) [43]. These barriers participate in the CNS homeostasis preventing the entrance of harmful substances from the circulation while transporting nutrients and metabolic compounds in and out of the brain.

The BCSFB is formed by the epithelial adjacent cells of the choroid plexus connected by tight junctions. The existence of microvilli on the apical surface and interdigitations on the basolateral surface, allow the exchange between the blood and the CSF [44].

Changes in BBB and BCSFB functions may be critical in age-related vulnerabilities and neurodegenerative diseases. Decrease in microvessel density, alterations of endothelial cells, including focal necrosis and reduced mitochondrial density, loosening of tight junctions and increased pinocytotic vesicles, together with decreased CSF production and turnover are among the most

described modifications in such conditions [43]. Although not clarified whether cerebral vascular dysfunction is a cause or a consequence of AD, it has been associated with disease progression and neuroinflammation [45,46]. Vascular lesions were found in the majority of cases of AD with late onset dementia and in nearly one half of those with early onset dementia [47]. Although leucocyte infiltration is not a predominant feature in the AD brain, increased T cell migration was observed in patients, albeit at low numbers [48]. Data also indicate a transmigration of monocytes into the CNS as a specific response to A β *in vitro*, similarly found in the brain of AD transgenic mice [49,50].

While BBB leakage [51] and impaired glial-vascular network involving astrocytic hypertrophic morphology and microvascular degeneration were found in aged rats [52], a BBB compromise was likewise observed in cerebral amyloid angiopathy [53]. Interestingly, the presence of microglial clusters, together with the increased expression of vascular endothelial growth factor (VEGF) in individuals with severe AD, was suggested to be related with vascular remodeling [54]. Interestingly, deposition of A β in the meningeal and intracerebral vessels was referred to be more age-related than AD-dependent [55]. However, we should not neglect that an increased causative effect of A β on the neurovascular injury may occur [56].

Endothelial cells were demonstrated to play a key role in BBB properties constituting its anatomic basis together with the basal membrane, although recent evidences indicate that such cells are intimately related with pericytes and astrocytes and interact with neurons, microglia and oligodendrocytes [57]. The endothelial cell monolayer is characterized by the absence of fenestrations, low pinocytotic vesicle number and elaborated tight and adherens junctions [58]. BBB dysfunction may derive from a simple transient opening of tight junctions to chronic barrier breakdown causing increased permeability. Acquisition of SASP phenotype in endothelial cells was shown to be associated with pathogenic miRNA signaling, (in Section 3.4), in particular miRNA(miR)-221, miR-155 and miR-21 [59,60]. In addition, the presence of miR-214 in endothelial cells that was shown to stimulate migration and angiogenesis in recipient cells was found decreased in senescent cells [61], thus justifying an impaired angiogenesis contributing to age-related cerebro-microvascular rarefaction and cognitive impairment. Also accounting to such dysregulation is the down-regulation of Dicer1 expression observed in aged endothelial cells [62].

2.2. Senescence of neurons and glial cells

Neurons are particularly vulnerable to age-related changes that affect their function and contribute to the onset of age-related neurodegenerative pathologies, namely AD and PD, although such association has been considered controversial [37]. More than the loss of neurons, their functional plasticity impairment by synaptic changes is considered to play a key role [63,64,65]. The extensive loss of axons during ageing was suggested to be implicated in the loss of connections and to trigger gradual cognitive decline, as well as sensorial and motor function failure [17]. The reduction in synaptic number and plasticity was shown to be associated to learning deficits in the ageing brain [65]. Such axon loss seems to not be a consequence of neuronal cell death, as axon loss really exceeds death of the corresponding cell bodies. As a matter of fact, no more than 10% neuron loss was found to occur by ageing [66]. The loss of neurites before cell death may derive from aberrant mitochondrial dynamics due to detrimental fission [67], although many other causes, including genetic and environmental factors may be considered as well [68]. A recent study using *in vivo* two-photon imaging in aged mice identified increased rates of axonal bouton dynamics and

synaptic instability associated with age-related cognitive impairment [69].

Recent evidence indicates that although the number of neocortical neurons do not differ between the aged 65–75 years individuals and the very old people with 94–105 years, there is a significant difference in the total number of neocortical glial cells between such age groups, mainly due to reductions in the number of oligodendrocytes [70]. The study is consistent with an age-related reduction in white matter volume previously observed in subjects at 80-year-old when compared to the age of 20 [71].

Astrocytes were shown to have a more inflammatory phenotype with age and to undergo a functional decline, both *in vivo* and *in vitro* studies, and to accumulate in aged brain [24,72]. Moreover, the inflammatory phenotype may disturb the integrity of the BBB since protein expression and distribution in endothelia are induced by astrocyte-derived signals, which also modulate BBB permeability and can be depleted in ageing [73,74]. The increased expression of the senescent biomarker p16^{INK4a} in senescent astrocytes was suggested to derive from the cumulative impact of stressors and to be associated to age-related susceptibility to sporadic AD [24]. Neuroglial aged cocultures have shown several senescence features that include progressive accumulation of SA- β -gal, increase in protein oxidative damage and in the proinflammatory cytokine IL-6, together with the down-regulation of miR-17, miR-19b, miR-20a, and miR-106b [75].

Studies point to a different distribution of microglia with ageing and to the existence of dysfunctional cells that may influence the functionality of the surrounding ones. Dopaminergic neurons seem to be particularly vulnerable to oxidative stress by the deregulated microglia [76] and the reduced clearance of A β peptide by the senescent microglia contributes to the pathogenesis of AD [77]. Smaller dendritic arbors and more elongated shape were observed in the aged microglia [78,79]. Dystrophic morphologies that include deramification, spheroid formation, shortened and twisted cytoplasmic processes, as well as partial or complete cytoplasmic fragmentation, were observed in the older brain [80,81]. However, such dystrophic morphology corresponds to only a microglia subtype located along with normal surveilling and ramified cells [82]. Indeed, microglia degeneration and loss of microglial neuroprotection, rather than activated microglia, was indicated to contribute to the onset of AD [83]. It has been hypothesized that during chronic mild neuroinflammation [84,85] the microglia phenotype switch from M2 (alternatively activated microglia) to M1 (classically activated) [86,87], but this inflammatory scenario may only occur in the early stages of the disease. Such features seem to disappear with AD progression [88] as microglia become less able to respond to A β peptide in elderly [89,90]. This picture is reinforced by the fact that nonsteroidal anti-inflammatory drugs (NSAIDs) were only successful when administered before the development of neurodegeneration [91]. When NSAIDs were administered in later stages of disease they even showed to be detrimental [92]. In this context it was demonstrated that microglia evidence an age-dependent decrease ability to phagocytose A β fibrils [93] and that older microglia internalize and redistribute less A β than young ones [90]. A recent study demonstrated that aged microglia in culture are less responsive to neurotoxins, such as bilirubin and A β , and evidenced less autophagic capacity and ability to migrate and phagocytose, reduced expression of toll-like receptor (TLR)-2, TLR-4, miR-124 and miR-155, together with increased SA- β -gal activity and miR-146a expression [94] (Figure 2). To note, however, that the poorly immunogenic properties of aged macrophages were suggested to be reversed by a proper environment or adequate stimulation [77]. Therapeutic approaches for the rejuvenation of microglia are discussed in section 5.2.

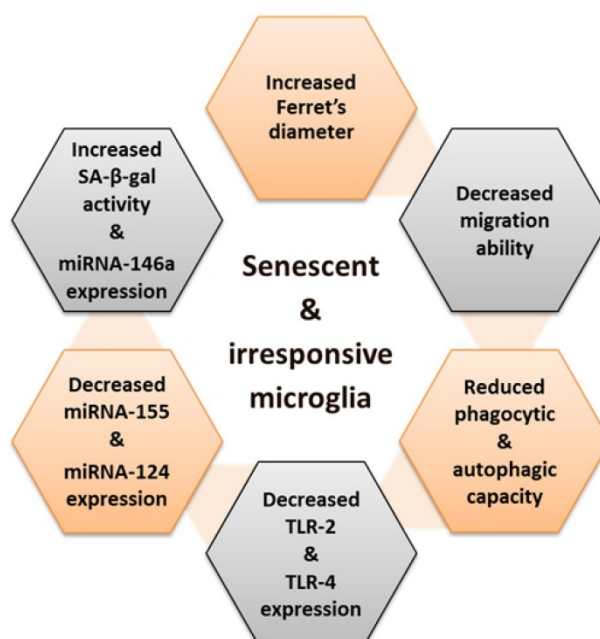


Figure 2. Hallmarks of senescence in aged cultures of microglia. Based on previous published work by Caldeira et al. [94].

3. Ageing promotes disease at the cellular level and local networks

Ageing is a progressive degenerative process tightly associated with chronic inflammation, derived from redox stress, mitochondrial damage, immunosenescence, and age-related diseases. In fact, although senescence and exhaustion may compromise immunity they seem to be distinct processes mediated by different signaling pathways, at least in T cells [95]. Immunosenescence is associated with decreased immune functions and cell proliferation in the presence of mitogenic or antigen stimulation in ageing immune cells [96]. The accumulation of age-related imperfections in cells and tissues may trigger inflammation in response to stressful conditions, where mitochondrial dysfunction can be both the cause or consequence [97]. This deterioration is a major risk factor for neurodegenerative diseases, and altered intercellular communication and inflammageing are important hallmarks [36]. Inflammageing results from prolonged exposure of the immune system to stimuli and is associated with age-related chronic disease, functional decline and frailty [98].

Lately, miRNAs are emerging as modulators of nuclear factor(NF)- κ B, mammalian target of rapamycin (mTOR), sirtuins, tumor growth factor(TGF)- β and Wnt signaling pathways, that may be related to inflammation (inflamma-miRs), cellular senescence (SA-miRs) and age-related diseases, including cancer-associated (onco-miRS) [99]. Another essential aspect is the disruption of the communication between neurons and the neighboring glial cells, as the neuronal fractalkine and its receptor CX3CR1 in microglia. Intriguingly, the natural reduction of fractalkine with age in hippocampus was shown to be associated with cognitive decline and its up-regulation may represent a novel therapeutic strategy [100].

New data indicate that formation and accumulation of protein aggregates may increase with ageing due to failure of protective pathways that include the unfolded protein response, the ubiquitin proteasome system and autophagy [101]. Autophagy is a recycling process essential for cell survival

and differentiation mediated by autophagosomes that engulf cytoplasmic contents and fuse with lysosomes, delivering their content that will be digested by *in situ* hydrolases [102]. Thus, autophagy represents an essential cytoprotective pathway that participates in the maintenance of homeostasis. Since accumulation of autophagic vacuoles is one of the pathologic hallmarks of degenerating neurons in AD, a causative connection between autophagy failure and neuronal death is believed to be present [103]. Interestingly, dysregulation of autophagy in neurons may lead to increased secretion of toxic proteins in exosomes favoring the spread of neurodegenerative diseases [104]. That's why a close relationship between autophagy and biogenesis and secretion of exosomes was recently suggested [105]. While autophagy declines with age, as observed by the decreased expression of key autophagy genes such as ATG5 and ATG7 in the brains of aged individuals [106], the release of exosomes is increased in senescent cells [107]. Therefore, how the two mechanisms are related and coordinated is still a matter of discussion.

3.1. Mitochondrial dysfunction

Mitochondrial dysfunction with accumulation of somatic mitochondrial DNA (mtDNA) mutations and decline in respiratory chain function is related with the ageing process [108]. Mitochondrial dysfunction has been associated with the increased generation of ROS, which affects replication and transcription of mtDNA causing mitochondria failure, and further damage of mtDNA and ROS production [109]. In aged individuals, mitochondria evidence a reduction in oxidative capacity, oxidative phosphorylation, ATP production and antioxidant defense [110]. Depolarized and damaged mitochondria are the substrate for mitophagy [111], a selective type of autophagy that eliminates damaged or unnecessary mitochondria [112]. In ageing-related diseases it is observed an accumulation of dysfunctional mitochondria by defects in mitophagy [113]. Autophagic removal of mitochondria is important for mitochondrial quality control. These two processes contribute to the preservation of energy and accumulation of damaged and aggregated proteins [114]. Therefore, many nutrients have been suggested to rejuvenate the aged mitochondria, including vitamins and other bio-energetic enhancers [115,116,117,118]. Alterations in the rate of mitochondrial fission and fusion may contribute to the decline in mitochondrial function during ageing [119] and mitophagy was found to depend on fission [120]. If mitochondria are not removed by mitophagy, the alterations on the mitochondrial permeability transition may determine the release of pro-apoptotic proteins and apoptosis. However, senescent cells are somehow resistant to apoptosis and both apoptosis and autophagy significantly decline with ageing [121,122]. In such conditions, increased mitochondrial biogenesis tends to overcome the bioenergetically dysfunctional mitochondria leading to an increase in mitochondrial mass [123]. Several interventions, such as caloric restriction, antioxidants, exercise and pyruvate, among others, have been proposed to influence mitochondrial biogenesis and to be therapeutically useful [124]. In a recent study, the serine protease Omi was identified as a novel regulator of mitochondrial biogenesis once it was shown to cleave the glycogen synthase kinase 3 β (GSK-3 β) which, if increased, triggers the degradation of the PPAR γ coactivator-1 α (PGC-1 α), causing an impairment of mitochondrial biogenesis and neurodegeneration [125]. GSK-3 β inhibitors were then suggested to have a key role in blocking inflammation [126,127] through the activation of autophagy [128].

3.2. Imbalance of cathepsins and of the CX3CL1/CX3CR1 signaling

There is increasing evidence that disturbance of the normal balance of the enzymatic activity of cathepsins and their extralysosomal localization are among the first consequences of ageing and age-associated neurodegenerative diseases [129]. The activation of the endosomal/lysosomal system was suggested to be associated with increased production of A β peptides [130]. The amyloid precursor protein (APP) is metabolized by secretory (β -site APP cleavage enzyme: BACE) and endosomal/lysosomal pathways (β -secretase activity of the cathepsins S, B and L) [131,132,133]. A β secretion revealed to be particularly linked to the ability of cathepsin S to generate A β from amyloidogenic fragments of β APP in the endosomal/lysosomal compartment [134]. Leakage of cathepsins into the cytoplasm subsequent to the disintegration of lysosomal membrane increase with ageing and if cathepsins B, H and L are relatively unstable [129], cathepsins D, E and S are stable and may degrade intracellular and extracellular proteins [135]. Up-regulation of the endosomal/lysosomal system was observed in damaged neurons of AD patients [130] and in activated microglia, where the secretion of cathepsin B and S has been indicated to induce neuronal death [136,137]. To consider that, apart of caspases, cathepsins are also associated with cell death regulation when released from the lysosome into the cytoplasm [138,139,140]. In particular, cathepsins have been shown to activate caspase-8 and to induce the cleavage of the pro-apoptotic Bcl-2 family member Bid, mitochondrial damage and subsequent caspase activation and apoptosis [141,142] (Figure 3).

Intriguingly, cathepsin S was recently shown to be involved in age-related inflammatory processes, AD and neuropathic pain [143,144,145,146], and its inhibition prevented the upregulation of proinflammatory factors in the brain cortex after traumatic brain injury in mice [147]. In addition, the release of cathepsin S from microglia was shown to be necessary for the cleavage of the transmembrane chemokine fractalkine or CX3CL1 from neurons [148,149], which further activates the pathway through the receptor CX3CR1 in microglia [150] (Figure 3). There is a lot of controversy on the role of CX3CL1-CX3CR1 axis on age-related neurodegenerative disorders and AD, as well as on the effects that CX3CR1 up- and down-regulation may have at the level of microglia migration and phagocytic ability towards A β . Some studies demonstrated that CX3CR1 expression was not altered in ageing or AD, while fractalkine gene expression was downregulated in the hippocampus, suggesting a loss of communication between neurons and microglia [151]. Deletion of CX3CR1 in a mouse model of AD increased the release of inflammatory cytokines, exacerbated neuronal dysfunction and potentiated cognitive deficits [152]. Lacking of the CX3CR1 receptor led to impairment of hippocampal cognitive function and synaptic plasticity [153]. However, other studies observed that both CX3CL1 and CX3CR1 decrease with advancing age [154,155] and that CX3CL1 signaling is deficient in AD brains and is down-regulated by A β [153]. In addition, plasma soluble CX3CL1 was higher in patients with mild to moderate AD than in the severe cases [156]. The issue is by no means clear and deserves further studies that demonstrate the role of such players in the process of ageing and in AD. Indeed, CX3CR1 deficiency showed to reduce A β deposition, as well as neuritic dystrophy, while enhancing microglia phagocytic ability [157], and CX3CL1 demonstrated to promote microglial phagocytosis of neuronal debris through the release of milk fat globule-EGF factor 8 (MFG-E8) [158], suggesting that benefits may be obtained by increasing fractalkine expression.

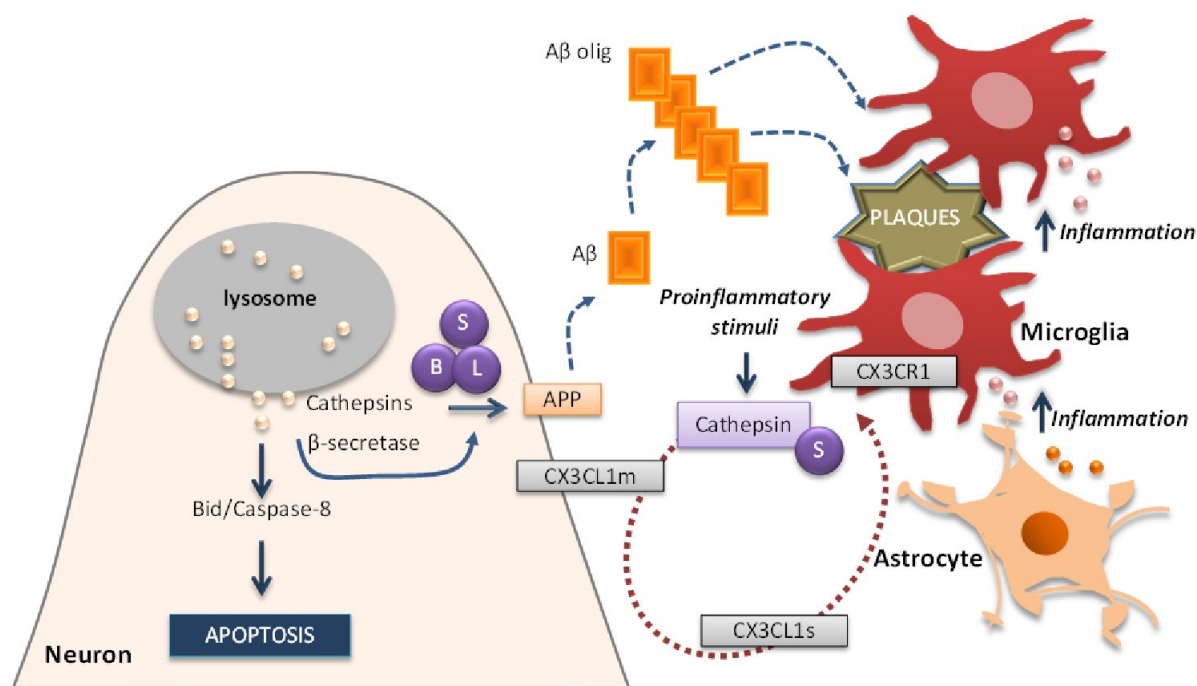


Figure 3. Role of cathepsins in the amyloid- β ($A\beta$) peptide processing and neuroinflammation in age-related neurodegenerative disorders, such as Alzheimer's disease. Activation of the endosomal/lysosomal system lead to the release of cathepsins into the cytosol. In such condition, and due to the β -secretase activity of the cathepsins S, B and L, it may occur increased production of amyloid- β ($A\beta$) peptides by enhanced metabolization of the amyloid precursor protein (APP). Disintegration of lysosomes and release of cathepsins are implicated in the proteolysis of caspase-8 and the Bcl-2 family member Bid, which subsequently activate the enzymatic signaling cascade leading to apoptosis. When $A\beta$ accumulates in the extracellular space of the brain it can aggregate into oligomers (olig) and form plaques. These plaques may disrupt cell-cell communication and activate microglia that trigger inflammation through the release of cytokines, which induce astrocyte reactivity and further release of pro-inflammatory cytokines. CX3CL1, or fractalkine, exists as a membrane-anchored (CX3CL1m) molecule, as well as in soluble form (CX3CL1s) when cleaved by cathepsin S originated from the activated microglia. The CX3CL1s then interacts with its receptor CX3CR1 in microglia sustaining the normal microglial activity. However, CX3CL1/CX3CR1 axis may play neuroprotective or neurotoxic roles depending on the brain injury.

In the normal brain, $A\beta$ is degraded within lysosomes, but in the AD brain it first accumulates within lysosomes and late endosomes, triggering the release of hydrolytic enzymes into the cytoplasm, followed by trafficking into autophagic vacuoles and neuronal death. Such degenerating neurons release the intracellular amyloid peptides into the extracellular space, which aggregate to form senile plaques that contain lysosomal proteases, including cathepsins [103]. Microglial cells were shown to aggregate around amyloid plaques in AD, to change their morphology and process motility [159], and to overexpress the potent pro-inflammatory cytokine IL-1 [160]. In ageing, the coverage of $A\beta$ plaques by microglia is reduced leading to enlarged protofibrillar $A\beta_{42}$ hotspots and

more severe neuritic dystrophy. When the *CX3CR1* gene was deleted it was found lower brain levels of A β 40 and A β 42, as well as reduced amyloid deposits, due to the increased microglial coverage and phagocytic ability together with a reduced neuritic dystrophy [161,162]. In what concerns astrocytes it is well known that stress and injury trigger the up-regulation of the expression of pro-inflammatory cytokines and chemokines, which are associated with the pathogenesis of AD where A β , S100B and IL-1 are associated with A β generation between astrocytes and neurons leading to sustained neuroinflammation [163] (Figure 2). In addition, enhanced cytokine levels were shown to increase BACE, APP and A β levels and stimulate amyloidogenic APP processing in astrocytes [164], contributing to the progression of the disease.

We may conclude that a better understanding of cathepsin imbalance during ageing and neurodegenerative diseases may contribute to the development of novel neuroprotective approaches. Additional studies are necessary to further define the role of CX3CL1/CX3CR1 signaling in the age-related susceptibilities to AD pathophysiology, while further elucidating microglial toxic or protective effects in order that therapeutic strategies may trigger microglia rejuvenation and neuroprotection by promoting A β clearance without inflammation.

3.3. Excessive accumulation of misfolded proteins

The accumulation of misfolded proteins is a recurring event during brain ageing and it was shown to be exacerbated in several neurodegenerative diseases. It has been suggested that protein accumulation may result from a dysfunction in the ubiquitin proteasome system [165]. Ageing is associated with progressive accumulation of structurally and functionally abnormal proteins, being oxidative stress a major contributor. Chronic oxidative stress is considered to lead to protein misfolding or unfolding, resulting in proteins unable to carry out their normal functions due to inefficient turnover by proteasomal and lysosomal pathways [166]. Such molecules that oligomerize and form aggregate structures are hallmarks of a range of neurodegenerative diseases including AD, PD and ALS, among others, all of them associated with ageing [167]. Proteostasis network decline in performance is also caused by mutations eventually leading to proteostasis collapse and cell death [168]. Selective proteotoxic toxicity is observed with specific proteins such as huntingtin and A β suggesting that neuronal cells may be maladapted to the stress of misfolded proteins. Enhanced autophagy and restoration of the control machinery capacity can have potential therapeutic benefits [169]. Damaged and misfolded proteins may bind to chaperones, which are highly conserved proteins that assist in the folding of proteins, thus leading to heat shock factors (HSFs) release [170]. HSFs are transcriptional activators of heat shock genes [171]. Cellular senescence was indicated to trigger the degradation of HSFs leading to an accelerated ageing [170]. In recent years, it was discovered that proteins in the cytosol can cross the lysosomal membrane, a process called chaperone-mediated autophagy, leading to the degradation of damaged or abnormal proteins. Such mechanism was shown to be impaired by ageing [172]. The deficiency seems to be related with alterations in the lipid composition of the lysosomal membrane mainly related with increased degradation of LAMP-2A.

3.4. Alteration of microRNAs in ageing and age-associated neurodegenerative diseases

miRNAs are short non-coding RNA molecules that intervene in gene regulation acting as

repressors, as well as activators, mainly at the post-transcriptional level [173,174]. Mature miRNAs are between 18–25 nucleotides in length. One miRNA can target multiple mRNAs, and one mRNA can be targeted by multiple, distinct miRNAs. Therefore miRNAs can significantly alter gene expression regulatory networks [175].

While not much has been published on miRNA secretion during the process of ageing, the potential of miRNAs to modulate ageing has attracted the interest of the scientific community, and their role in controlling ageing process has been uncovered recently [175,176]. Their differential expression during ageing in the mouse brain suggests that multiple gene regulatory relationships are affected and that insulin signaling pathway is also disturbed [177].

When evaluated in human serum during the ageing process it was observed that five miRNAs were down-regulated (miR-29b, miR-106b, miR-130b, miR-142-5p, and miR-340) and three were up-regulated (miR-92a, miR-222, and miR-375) [178]. It has been claimed that the loss of miRNA synthesis in adults reduces lifespan and results in rapid ageing [179]. However, circulating miR-146a and miR-21 were shown to be enhanced by ageing in mice, and in age-related diseases (including AD), respectively [180,181], while miR-155 in blood was found inversely correlated with age in patients with coronary artery disease [182] and in a cohort of Whites and African Americans between the ages of 30–64 [183]. In our model of aged microglia, both miR-124 and miR-155 expression were decreased when compared with young/reactive microglia, but expression of miR-146a was clearly increased [94]. Such up-regulation may occur to prevent excessive inflammatory response considering that IL-6 and IL-8 are targets of miR-146a [176]. Among the several hallmarks of ageing and the associated miRNAs we may refer to miR-146a as correlated with mitochondrial dysfunction and “inflammageing”, miR-34a with mitochondrial dysfunction and telomere attrition, miR-155 with the loss of telomeres and “inflammageing”, miR-21 with altered DNA damage response and “inflammageing”, miR-371, miR-29, miR-499 and Let-7 with stem cell exhaustion, and miR-17, miR-19b, miR-20a and miR-106a cluster with the regulation of cellular senescence [2]. There is now evidence that miR-155, miR-21, miR-124 and miR-146a, known as the inflamma-miR group, are involved in the process of “inflammageing” due to increased inflammation on a background of reduced immune capacity [2,184,185]. Despite this small number of inflamma-miRs indicated as fine tuners of inflammation likely associated with the cellular SASP acquisition hallmark, the molecular mechanisms of their action have proved to be highly complex.

When the AD hippocampus was compared with the adult brain it was observed that miR-9 and miR-128 were upregulated [186]. Similarly, miR-34a was highly expressed in the cerebral cortex of APP^{swe}/PSΔ^{E9} mice, a murine model of AD, and evidenced to be inversely correlated with the Bcl2 protein [23]. In addition, miRNA-9, miRNA-125b and miRNA-146a were found to be up-regulated in AD patients shortly after death [187]. In contrast, neuronal miR-107 was found decreased, even in the early stages [22], together with the miR-29a/b-1 cluster, the later suggested to be related with increased BACE and A β levels in sporadic AD [188]. Our preliminary results have shown increased expression of miR-155 in the cortical and hippocampal brain regions of 3-month 3x-Tg-AD mice (unpublished data), when there is increasead intracellular A β , but low levels of extracellular A β [189,190]. Upregulation of miR-146a is commonly observed in AD, but also in prion disease, although it was not clarified whether it represents the cause or the consequence of the disease [191]. A recent study identified 12 potential circulating miRNAs that may be used in AD diagnosis [165], two of which (miR-103 and miR-107) already suggested to be implicated in the disease [177].

Considering the potentialities of miRNAs as biomarkers and the possibility of their modulation

as neuroprotective therapeutic approaches, further investigations are required. Overall, little is understood about the role of miRNAs in AD, and even less when age stratification is considered.

4. Role of exosomes and ectosomes in neurodegeneration, clinical diagnosis and drug delivery

Almost all living cells release extracellular vesicles (EVs) that facilitate intercellular communication by carrying proteins, lipids, mRNAs and miRNAs [192,193,194,195] (Figure 1). EVs are categorized into exosomes and ectosomes or microvesicles (MVs), based on their mechanisms of biogenesis and biophysical properties, such as size and surface protein markers [196]. Exosomes are homogenous small particles with 30–100 nm in size that derive from endosomal compartments, called multivesicular bodies (MVBs), after fusion with the plasma membrane [197]. Ectosomes constitute a larger and more heterogeneous population of EVs, with 100–1000 nm in size. They are produced directly through the outward budding and fission of membrane vesicles from the plasma membrane [198,199].

Recently, it was shown that some miRNAs are specifically sorted in exosomes, whereas others are retained in cells, thus signifying the existence of mechanisms controlling the sorting into exosomes [200]. Ectosomes mainly contain mRNAs and comprise less miRNAs than exosomes [27]. Despite such differences it is difficult to discriminate exosomes from ectosomes upon release from cells and their functions are believed to be largely analogous [201]. They are able to travel long distances to their target cells in which the cargo is delivered via ligand-receptor interaction, fusion with plasma membrane, or internalization by endocytosis [202] (Figure 1). Additionally, a latest study showed the presence and differential abundance of long non-coding RNAs (lncRNAs) molecules longer than 200 nucleotides [203] in secreted exosomes [204], indicating a selective packing, as also described for mRNAs and miRNAs [195,205].

4.1. EVs as accelerators of ageing and mediators of neurodegenerative diseases

The pathophysiological significance of extracellular vesicles is beginning to be defined in diseases, including neurodegenerative diseases. It was shown that exosomes are secreted by neural cells (including neurons, astrocytes, oligodendrocytes and microglia) under both normal and pathological conditions [206]. Microglial exosomal vesicles were shown to express the aminopeptidase CD13 and to contain major histocompatibility (MHC) class II molecules and cathepsin S [207]. In addition, the release of MVs by microglia represents a secretory pathway for IL-1 β , thus influencing neuronal and non-neuronal cells. ATP, the receptor P2X₇ and acid sphingomyelinase regulate the shedding of MVs from microglia [208].

Exosomes carrying α -synuclein (α Syn) oligomers were shown to induce more toxicity than free α Syn oligomers [209] and were suggested to spread the disease to neighboring cells [206]. The fact that exosomal miRNAs may be obtained from frozen postmortem brain tissue [210] enlarge the possibilities of being used as biomarkers of neurodegenerative diseases, such as autism, schizophrenia and bipolar disorder [210,211,212,213]. Lately, it was demonstrated that Tau, a microtubule-associated protein that aggregates in neurodegenerative diseases, is rather secreted in ectosomes than in exosomes [214]. Back studies have revealed that the A β that accumulates in AD is released in association with exosomes and that amyloid plaques are enriched in exosomal proteins [215]. Neuron-derived exosomes are suggested to assemble A β that is internalized by microglia, thus

participating in the clearance of extracellular A β [216]. A recent study evidenced that neuronal exosomes, but not glial exosomes, are involved in A β clearance and that the reduction in exosome release is age-related [217]. However, it was also demonstrated that MVs from AD patients are toxic for cultured neurons due to the delivery of neurotoxic A β species generated through the intervention of membrane lipid components of the MVs [218]. Furthermore, one should also consider that MVs may seed and feed A β assembly with neurotoxic properties while favoring cell-to-cell spreading [219]. It was lately suggested that exosomes released by activated neurons sensitize microglia for phagocytic clearance of inappropriate synapses, promoting the synaptic pruning by microglia [220]. Microglia internalize oligodendroglia-derived exosomes by macropinocytosis, participating in the degradation of oligodendroglial membrane in an immunologically silent manner, i.e. without microglia activation and in a subpopulation of MHCII-negative cells [221].

Exosomes derived from astrocytes and motor neurons revealed to play a key role in ALS disease based on studies demonstrating the efficient transfer of mutant copper-zinc superoxide dismutase 1 (SOD1) and misfolded SOD1 to motor neurons, causing cell death and the spread of the disease [222,223]. Overexpression of mutated SOD1 is included in 5–10% of ALS cases from familiar origin, although malformed SOD1 was also found in sporadic ALS samples [224] and the propagation and transmission of misfolded SOD1 considered the cause of most, if not all, types of ALS [223]. Therefore, exosomes are now suggested as targets to modulate ALS disease [222].

There are only a few studies on cellular senescence and associated alterations in exosome biogenesis and cargo. In a previous study using an accelerated form of senescence by irradiation in human prostate cancer cells it was observed that such treatment produced replicative cellular senescence, inducing a p53-dependent increase in the biogenesis of exosome-like vesicles [107]. Although the release of exosomes was shown to increase during accelerated and replicative cellular senescence [107] intriguing results were obtained in models of AD. Impairment of endocytic transport was suggested to be responsible for the decreased number of exosomes found in the CSF of APP transgenic mice at 23 months of age when compared to that found at 2 months-old [217]. This fact would explain the deficient clearance of A β and tau that are known to be associated to EVs [219]. Epigenomic alterations during ageing are suggested to increase exosome secretion together with an altered expression of circulating miRNAs, exosome proteins and cytokines, which will influence cell-cell communication mediated by exosomes [225]. Also reinforcing a different composition by ageing is a recent study evidencing that exosomes isolated from sera of old subjects contain higher levels of IL-6 and IL-12 mRNA than those from young individuals, which further indicates that ageing may influence the mRNA delivery by exosomes [226]. The latest concept that exosomes may be used as diagnostic biomarkers due to their different signature cargo [227] opens new fields of investigation, in particular in their role in ageing-related vulnerabilities.

4.2. EVs as biomarkers of disease and drug delivery

It is considered that the molecular content of exosomes reproduces the cell of origin and its status. For instance, misfolded proteins associated with ALS, PD, AD and neurodegenerative tauopathies are processed by MVBs and found in exosomes [228]. Identification of elevated levels of total and phosphorylated tau, as well as of A β , were observed in blood exosomes from patients with AD and fronto-temporal dementia and suggested to be potential biomarkers for the staging of sporadic AD, as well as predictors of the disease [229]. Similarly, exosomal miRNA signature may

additionally constitute a suitable screening tool in serum from AD patients [230], based on the elevated content of miR-193b in blood and CSF-derived exosomes [231] of the APP/PS1 double-transgenic AD mice model using miRNA array technology [231]. A comprehensive revision on the potentialities of miRNA profiling as biomarkers in neurodegenerative diseases was recently published [232].

The potential application of exosomes as biomarkers and their biomedical utility reside on the higher specificity and sensitivity they have over those in serum or urine, as a consequence of their elevated stability and facility to be isolated from biofluids, including saliva [233]. Exosomal small RNAs, including miRNAs, are protected against RNase A treatment, but their profile and concentration may differ between cells, plasma or serum and exosomes in blood [234]. The potential value of exosomes in clinical diagnostics has just begun to be explored and larger cohort studies are needed to achieve their validation as biomarkers.

Exosomes evidence several advantages as delivery vehicles due to long half-life in circulation, low immunogenicity and ability to cross the BBB, while may be genetically engineered to package mRNAs, siRNAs, proteins and drugs [235]. Exosomes, instead of synthetic liposomes do not have the limitation of the lipid membrane toxicity, but there are no ideal, scalable and cost effective methods to obtain a high-yielding source of purified exosomes [236]. If such problem is surpassed we are cautiously optimistic that engineered exosomes may be useful in delaying or preventing age-related neurodegenerative diseases, thus transforming health care.

5. Therapeutic opportunities

Potential strategies for mitigating the deleterious effects of senescent cells and fight ageing include transplants of neural stem cells or more specialized differentiated cells and rejuvenation approaches [237,238]. Direct reprogramming of somatic cells into induced pluripotent stem cells (iPSCs) even from senescent and centenarian cells has become a powerful technique in tissue replacement therapies [239]. Methods require the passage of the old cell through human embryonic stem cells (hES) and iPSCs, but the aged phenotype can also be reverted back to a youthful one by age reprogramming approaches [240]. Age-related neurodegenerative diseases represent a terrible societal burden and cellular replacement of lost cells and/or enrichment environment in glial cells may delay neurodegeneration and provide beneficial effects by both strategies [241]. To avoid rejection when using interindividual transplantations the usage of a person's own stem cells for transplantation may have additional advantages and hES an added value due to their multipotent properties and reduced reactivity toward immune cells [242]. Clinical application of such therapeutic approaches requires considering the pathological/inflammatory neuronal environments, as well as the safety and treatment benefits [243]. Significant advances and promising results have already been achieved in ALS clinical trials [244] and it is conceivable that future clinical trials may be approved for other neurodegenerative diseases.

5.1. Rejuvenation of senescent cells

The rejuvenation of cell senescence through the reversal/inhibition of cellular ageing may extend the lifespan. Main approaches use genetic or pharmacological approaches and nutritional alterations. The recent discovery of iPSCs obtained from the patient cells (e.g. fibroblasts,

keratinocytes) has opened the possibility of autologous regenerative medicine, although it was shown to favor the formation of tumors [245]. Another way to prolong lifespan uses rapamycin, an inhibitor of mTOR pathway that showed to inhibit cellular senescence [246]. Rapamycin, a drug approved by the Federal Drug Administration (FDA) used to prevent organ rejection after kidney transplantation, was the first pharmacological compound to increase longevity in mammalian species [247]. However, rapamycin also triggers inflammatory side effects [248,249]. Recently, the anti-senescence effect of statins was suggested based on its ability to inhibit telomere shortening [250]. Rejuvenation of antioxidant system that was shown to be decreased in the CNS of aged rats was achieved by grape seed extract with ability to decrease the free radical-induced lipid peroxidation [251]. Age-related increase in lipid peroxidation and declined antioxidants may eventually be counterbalanced by the consumption of vitamins B, C, D and E, flavonoids, polyunsaturated omega-3 fatty acids and fish [252,253,254], which seem to be important in the prevention of cognitive decline, as well as of dementia and AD [255]. Energetic insufficiency and depleted energy reserve in brain tissue are implicated in several neurodegenerative diseases [256], and mitochondrial dysfunction has been associated to age-related diseases, such as AD [257]. Therefore, several approaches have been indicated to rejuvenate mitochondria [258,259] and mitochondrial properties showed to return to a pre-programmed state in iPSCs from fibroblasts [260]. Nevertheless, it was demonstrated that iPSCs kept for prolonged periods *in vitro* present accumulation of defective mitochondria [261]. Proposed therapeutic approaches for the rejuvenation of senescent microglia include anti-inflammatory (e.g. minocycline, IL1 receptor antagonist), anti-oxidant (e.g. flavonoids, retinoid/carotenoid class components, vitamins E and D3) and measures to stimulate autophagy, a process slowed by lipofuscin accumulation [238]. A recent study has demonstrated that the inhibition of the COX/PGE2/EP2 immune pathway restores microglial chemotaxis and A β clearance and suppresses overinflammation showing ability to restore healthy microglial function and to prevent AD progression [262]. Johansson et al. [251] observed an increased EP2 mRNA in aged macrophages in response to A β , not observed in young cells.

5.2. Cell replacement therapy

Cell replacement therapy has been proposed as a therapy for age-related microglia senescence and motor neuron diseases, such as ALS. Replenishment of old microglia by bone marrow-derived cells demonstrated that these cells acquire features of microglia within the CNS parenchyma and can be used as therapy [263]. Nevertheless, monocyte-derived macrophages revealed to be functionally distinct from the resident microglia assisting them [264,265]. It was lately successfully accomplished the intranasal delivery of microglia/macrophages and mesenchymal stem cells to the brain of transgenic PD and AD mouse models [266]. However, such approaches are limited by the occurrence of strong inflammatory response and invasion of peripheral monocytes/macrophages expressing MHC class II, a M1 microglial marker of activated cell [267].

ALS is an incurable adult-onset neurodegenerative disease, difficult to diagnose, and with limited survival expectancy due to the lack of effective treatments. Therapeutic difficulties arise from the still not clarified molecular mechanisms of motor neuron loss in ALS, which lead to muscle paralysis and death [268]. Thus, there is a growing interest in the intraspinal transplantation of neural stem cells to replace lost motor neurons. In a phase 1 FDA-approved clinical trial [269], intraspinal transplantation of neural stem cells (NSI-566RSC HSSC line) was considered feasible and safe, but

clinical benefit was somehow reduced. Even when an increased number of human neural stem cell was transplanted and reduced immunosuppression used, the benefits were still reduced [270]. To that it may account the excessive reactivity of local astrocytes that showed an aberrant phenotype when isolated from the spinal cord [271] and the cortical brain region (our unpublished data). Intriguingly, astrocytes from both the sporadic and familiar forms of ALS have shown toxicity toward motor neurons [272]. Most interesting, the intraspinal transplantation of glial-rich neural progenitors derived from human iPSCs in the transgenic mouse model of ALS improved the lifespan of the animal [273]. Lately it was demonstrated that human iPSCs can be used to efficiently derive neural progenitor cells, which after transplantation were shown to integrate into the rodent spinal cord and mature to astroglia [274]. It has been hypothesized that the transplantation of the stem cell secretome (e.g. extracellular membrane vesicles) may originate a more efficient outcome than the current-stem cell therapies due to the release of immunomodulatory or neurotrophic paracrine factors [275].

AD is the most common neurodegenerative disease in the elderly being responsible for about 50–80% of dementia cases. Patients evidence a chronic and progressive decline in memory that reduces a person's ability to perform everyday activities. Current therapies only alleviate symptoms without stopping neural degeneration or recovery [276]. Therefore, induction of neurogenesis, introduction of neural stem cells and microenvironmental alterations under AD pathology have been attempted in the mice model but not performed in AD patients [277]. A recent work evidenced that neural stem cell transplantation in the APP/PS1 transgenic mice enhanced mitochondria biogenesis and rescued cognitive deficits. However, although improving cognitive function, the neural stem cell transplantation revealed to not alter A β pathology [278]. Moreover, adipose-derived mesenchymal stem cell transplantation in this same model also showed to promote neurogenesis [279].

6. Conclusions

Ageing, characterized by a progressive loss of cell functionality, is nowadays the subject of a high number of studies intending to define the major hallmarks and the underlying mechanisms with the final goal of developing target-driven medicines and strategies that can improve/extend human health in the elderly. Age-related deterioration is the primary risk for major human pathologies, including neurodegenerative diseases. The number of people over 60 has doubled since 1980 and is predicted to outnumber children under the age of 5 within the next five years (World Health Organization). The prevalence of AD doubles for every 5-year interval beyond age 65 [280] and is responsible for about 50–80% of dementia cases, representing a major public health issue and a challenge to the health care system, with a tremendous impact at both the individual and the societal levels [281,282,283,284]. However, specific markers of such deterioration are missing and complexity arise from the existence of different categories: primary hallmarks (e.g. DNA damage), antagonistic hallmarks (e.g. senescence that protects from cancer at low levels but that promotes ageing when in excess) and integrative hallmarks (e.g. stem cell exhaustion and altered intercellular communication) [36]. Although the progression of neural stem cell niche demise may contribute to the ageing of the brain [285], it is not clear how cells are affected. It is suggested that they are not receiving the appropriate signals to maintain proliferation and neurogenesis [286]. Therefore, stimulation may become the focus of strategies to compensate neurogenesis decrease [287,288]. However, one should also consider that it may lead to the exhaustion of the neural stem cell pool.

It is therefore important at this time to define and identify specific criteria, as well as

appropriate standardized measures and models of frailty and ageing [289,290,291], to test novel interventions at promoting healthier ageing with the final aim of decreasing the prevalence of neurodegenerative diseases in old people. To improve the quality of European ageing research a coordinated interdisciplinary network was recently formed to reach consensus on ways to test preclinical interventions in ageing mice (BMBS COST Action BM1402), since it became the favorite mammalian model [292].

In this paper, we summarized current data on candidate hallmarks and contribution to the process of neural cell ageing (Table 1), with a particular focus on cell-cell communication deregulation, altered miRNA profiling and role of extracellular vesicles. More attention needs to be paid to the interconnectedness between such candidate hallmarks and their relative contributions to ageing. These are important issues to consider when tentative medicines are to be tested in improving healthspan. In addition, compelling evidence indicates that the speed of the ageing process may depend from genetic and epigenetic changes experienced among individuals during life and possibly will require, in the future, different interventions to enable life quality and reverse progress of neurodegenerative diseases in each person. Future research in these areas is sorely needed to be translated into health promotion practice for older people.

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Conflict of interest

The author declares no conflicts of interest in this paper.

References

1. Emde A, Hornstein E (2014) miRNAs at the interface of cellular stress and disease. *Embo J* 33: 1428-1437.
2. Harries LW (2014) MicroRNAs as Mediators of the Ageing Process. *Genes (Basel)* 5: 656-670.
3. Chen Z, Trotman LC, Shaffer D, et al. (2005) Crucial role of p53-dependent cellular senescence in suppression of Pten-deficient tumorigenesis. *Nature* 436: 725-730.
4. Galluzzi L, Bravo-San Pedro JM, Kroemer G (2014) Organelle-specific initiation of cell death. *Nat Cell Biol* 16: 728-736.
5. Hung CW, Chen YC, Hsieh WL, et al. (2010) Ageing and neurodegenerative diseases. *Ageing Res Rev* 9 Suppl 1: S36-46.
6. Rodier F, Campisi J (2011) Four faces of cellular senescence. *J Cell Biol* 192: 547-556.
7. Campisi J, Andersen JK, Kapahi P, et al. (2011) Cellular senescence: a link between cancer and age-related degenerative disease? *Semin Cancer Biol* 21: 354-359.
8. Coppé JP, Desprez PY, Krtolica A, et al. (2010) The senescence-associated secretory phenotype: the dark side of tumor suppression. *Annu Rev Pathol* 5: 99-118.

9. Lindsay J, Laurin D, Verreault R, et al. (2002) Risk factors for Alzheimer's disease: a prospective analysis from the Canadian Study of Health and Aging. *Am J Epidemiol* 156: 445-453.
10. Katz MJ, Lipton RB, Hall CB, et al. (2012) Age-specific and sex-specific prevalence and incidence of mild cognitive impairment, dementia, and Alzheimer dementia in blacks and whites: a report from the Einstein Aging Study. *Alzheimer Dis Assoc Disord* 26: 335-343.
11. Hindle JV (2010) Ageing, neurodegeneration and Parkinson's disease. *Age Ageing* 39: 156-161.
12. Van Den Eeden SK, Tanner CM, Bernstein AL, et al. (2003) Incidence of Parkinson's disease: variation by age, gender, and race/ethnicity. *Am J Epidemiol* 157: 1015-1022.
13. Mehta P, Antao V, Kaye W, et al. (2014) Prevalence of amyotrophic lateral sclerosis - United States, 2010-2011. *MMWR Surveill Summ* 63 Suppl 7: 1-14.
14. Kurtzke JF (1982) Epidemiology of amyotrophic lateral sclerosis. *Adv Neurol* 36: 281-302.
15. Choi JY, Morris JC, Hsu CY (1998) Aging and cerebrovascular disease. *Neurol Clin* 16: 687-711.
16. Popa-Wagner A, Buga AM, Turner RC, et al. (2012) Cerebrovascular disorders: role of aging. *J Aging Res* 2012: 128146.
17. Adalbert R, Coleman MP (2012) Axon pathology in age-related neurodegenerative disorders. *Neuropathol Appl Neurobiol*.
18. Zlokovic BV (2011) Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 12: 723-738.
19. Chen RL, Kassem NA, Redzic ZB, et al. (2009) Age-related changes in choroid plexus and blood-cerebrospinal fluid barrier function in the sheep. *Exp Gerontol* 44: 289-296.
20. Mao P, Reddy PH (2011) Aging and amyloid beta-induced oxidative DNA damage and mitochondrial dysfunction in Alzheimer's disease: implications for early intervention and therapeutics. *Biochim Biophys Acta* 1812: 1359-1370.
21. Castello MA, Soriano S (2014) On the origin of Alzheimer's disease. Trials and tribulations of the amyloid hypothesis. *Ageing Res Rev* 13: 10-12.
22. Herbig U, Ferreira M, Condel L, et al. (2006) Cellular senescence in aging primates. *Science* 311: 1257.
23. Jeyapalan JC, Sedivy JM (2008) Cellular senescence and organismal aging. *Mech Ageing Dev* 129: 467-474.
24. Bhat R, Crowe EP, Bitto A, et al. (2012) Astrocyte senescence as a component of Alzheimer's disease. *PLoS One* 7: e45069.
25. Dimri GP, Lee X, Basile G, et al. (1995) A biomarker that identifies senescent human cells in culture and in aging skin in vivo. *Proc Natl Acad Sci U S A* 92: 9363-9367.
26. Sadallah S, Eken C, Schifferli JA (2011) Ectosomes as modulators of inflammation and immunity. *Clin Exp Immunol* 163: 26-32.
27. Weilner S, Schraml E, Redl H, et al. (2013) Secretion of microvesicular miRNAs in cellular and organismal aging. *Exp Gerontol* 48: 626-633.
28. Hayflick L (1965) The Limited in Vitro Lifetime of Human Diploid Cell Strains. *Exp Cell Res* 37: 614-636.
29. Lee M, Lee JS (2014) Exploiting tumor cell senescence in anticancer therapy. *BMB Rep* 47: 51-59.

30. Campisi J, di Fagagna FD (2007) Cellular senescence: when bad things happen to good cells. *Nat Rev Mol Cell Biol* 8: 729-740.
31. Kurz DJ, Decary S, Hong Y, et al. (2000) Senescence-associated (beta)-galactosidase reflects an increase in lysosomal mass during replicative ageing of human endothelial cells. *J Cell Sci* 113 (Pt 20): 3613-3622.
32. Lee BY, Han JA, Im JS, et al. (2006) Senescence-associated beta-galactosidase is lysosomal beta-galactosidase. *Aging Cell* 5: 187-195.
33. Campo-Trapero J, Cano-Sanchez J, Palacios-Sanchez B, et al. (2008) Cellular senescence in oral cancer and precancer and treatment implications: a review. *Acta Oncol* 47: 1464-1474.
34. Collado M, Gil J, Efeyan A, et al. (2005) Tumour biology: senescence in premalignant tumours. *Nature* 436: 642.
35. Byun HO, Han NK, Lee HJ, et al. (2009) Cathepsin D and eukaryotic translation elongation factor 1 as promising markers of cellular senescence. *Cancer Res* 69: 4638-4647.
36. Lopez-Otin C, Blasco MA, Partridge L, et al. (2013) The hallmarks of aging. *Cell* 153: 1194-1217.
37. Morrison JH, Hof PR (1997) Life and death of neurons in the aging brain. *Science* 278: 412-419.
38. Ren JL, Pan JS, Lu YP, et al. (2009) Inflammatory signaling and cellular senescence. *Cell Signal* 21: 378-383.
39. Freund A, Orjalo AV, Desprez PY, et al. (2010) Inflammatory networks during cellular senescence: causes and consequences. *Trends Mol Med* 16: 238-246.
40. Molofsky AV, Slutsky SG, Joseph NM, et al. (2006) Increasing p16INK4a expression decreases forebrain progenitors and neurogenesis during ageing. *Nature* 443: 448-452.
41. Krouwer VJ, Hekking LH, Langelaar-Makkinje M, et al. (2012) Endothelial cell senescence is associated with disrupted cell-cell junctions and increased monolayer permeability. *Vasc Cell* 4: 12.
42. Abbott NJ, Friedman A (2012) Overview and introduction: the blood-brain barrier in health and disease. *Epilepsia* 53 Suppl 6: 1-6.
43. Marques F, Sousa JC, Sousa N, et al. (2013) Blood-brain-barriers in aging and in Alzheimer's disease. *Mol Neurodegener* 8: 38.
44. Redzic ZB (2013) Studies on the human choroid plexus in vitro. *Fluids Barriers CNS* 10: 10.
45. Klohs J, Rudin M, Shimshek DR, et al. (2014) Imaging of cerebrovascular pathology in animal models of Alzheimer's disease. *Front Aging Neurosci* 6: 32.
46. Jellinger KA (2002) Alzheimer disease and cerebrovascular pathology: an update. *J Neural Transm* 109: 813-836.
47. Carotenuto A, Rea R, Colucci L, et al. (2012) Late and early onset dementia: what is the role of vascular factors? A retrospective study. *J Neurol Sci* 322: 170-175.
48. Togo T, Akiyama H, Iseki E, et al. (2002) Occurrence of T cells in the brain of Alzheimer's disease and other neurological diseases. *J Neuroimmunol* 124: 83-92.
49. Lebson L, Nash K, Kamath S, et al. (2010) Trafficking CD11b-positive blood cells deliver therapeutic genes to the brain of amyloid-depositing transgenic mice. *J Neurosci* 30: 9651-9658.
50. Stalder AK, Ermini F, Bondolfi L, et al. (2005) Invasion of hematopoietic cells into the brain of amyloid precursor protein transgenic mice. *J Neurosci* 25: 11125-11132.

51. Enciu AM, Gherghiceanu M, Popescu BO (2013) Triggers and effectors of oxidative stress at blood-brain barrier level: relevance for brain ageing and neurodegeneration. *Oxid Med Cell Longev* 2013: 297512.
52. Zhang R, Kadar T, Sirimanne E, et al. (2012) Age-related memory decline is associated with vascular and microglial degeneration in aged rats. *Behav Brain Res* 235: 210-217.
53. Hawkes CA, Hartig W, Kacza J, et al. (2011) Perivascular drainage of solutes is impaired in the ageing mouse brain and in the presence of cerebral amyloid angiopathy. *Acta Neuropathol* 121: 431-443.
54. Jantaratnotai N, Schwab C, Ryu JK, et al. (2010) Converging perturbed microvasculature and microglial clusters characterize Alzheimer disease brain. *Curr Alzheimer Res* 7: 625-636.
55. Kovari E, Herrmann FR, Hof PR, et al. (2013) The relationship between cerebral amyloid angiopathy and cortical microinfarcts in brain ageing and Alzheimer's disease. *Neuropathol Appl Neurobiol* 39: 498-509.
56. Hartz AM, Bauer B, Soldner EL, et al. (2012) Amyloid-beta contributes to blood-brain barrier leakage in transgenic human amyloid precursor protein mice and in humans with cerebral amyloid angiopathy. *Stroke* 43: 514-523.
57. Cardoso FL, Brites D, Brito MA (2010) Looking at the blood-brain barrier: molecular anatomy and possible investigation approaches. *Brain Res Rev* 64: 328-363.
58. Sá-Pereira I, Brites D, Brito MA (2012) Neurovascular unit: a focus on pericytes. *Mol Neurobiol* 45: 327-347.
59. Tsai PC, Liao YC, Wang YS, et al. (2013) Serum microRNA-21 and microRNA-221 as Potential Biomarkers for Cerebrovascular Disease. *J Vasc Res* 50: 346-354.
60. Staszal T, Zapala B, Polus A, et al. (2011) Role of microRNAs in endothelial cell pathophysiology. *Pol Arch Med Wewn* 121: 361-366.
61. van Balkom BW, de Jong OG, Smits M, et al. (2013) Endothelial cells require miR-214 to secrete exosomes that suppress senescence and induce angiogenesis in human and mouse endothelial cells. *Blood* 121: 3997-4006, S3991-3915.
62. Ungvari Z, Tucsek Z, Sosnowska D, et al. (2013) Aging-induced dysregulation of dicer1-dependent microRNA expression impairs angiogenic capacity of rat cerebrovascular endothelial cells. *J Gerontol A Biol Sci Med Sci* 68: 877-891.
63. Burke SN, Barnes CA (2006) Neural plasticity in the ageing brain. *Nat Rev Neurosci* 7: 30-40.
64. Morrison JH, Baxter MG (2012) The ageing cortical synapse: hallmarks and implications for cognitive decline. *Nat Rev Neurosci* 13: 240-250.
65. Burke SN, Barnes CA (2010) Senescent synapses and hippocampal circuit dynamics. *Trends Neurosci* 33: 153-161.
66. Pakkenberg B, Gundersen HJ (1997) Neocortical neuron number in humans: effect of sex and age. *J Comp Neurol* 384: 312-320.
67. Arnold B, Cassady SJ, VanLaar VS, et al. (2011) Integrating multiple aspects of mitochondrial dynamics in neurons: age-related differences and dynamic changes in a chronic rotenone model. *Neurobiol Dis* 41: 189-200.
68. Reitz C, Mayeux R (2014) Alzheimer disease: epidemiology, diagnostic criteria, risk factors and biomarkers. *Biochem Pharmacol* 88: 640-651.
69. Grillo FW, Song S, Teles-Grilo Ruivo LM, et al. (2013) Increased axonal bouton dynamics in the aging mouse cortex. *Proc Natl Acad Sci U S A* 110: E1514-1523.

70. Fabricius K, Jacobsen JS, Pakkenberg B (2013) Effect of age on neocortical brain cells in 90+ year old human females--a cell counting study. *Neurobiol Aging* 34: 91-99.
71. Marnier L, Nyengaard JR, Tang Y, et al. (2003) Marked loss of myelinated nerve fibers in the human brain with age. *J Comp Neurol* 462: 144-152.
72. Pertusa M, Garcia-Matas S, Rodriguez-Farre E, et al. (2007) Astrocytes aged in vitro show a decreased neuroprotective capacity. *J Neurochem* 101: 794-805.
73. Zlokovic BV (2008) The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron* 57: 178-201.
74. Popescu BO, Toescu EC, Popescu LM, et al. (2009) Blood-brain barrier alterations in ageing and dementia. *J Neurol Sci* 283: 99-106.
75. Bigagli E, Luceri C, Scartabelli T, et al. (2015) Long-term Neuroglial Cocultures as a Brain Aging Model: Hallmarks of Senescence, MicroRNA Expression Profiles, and Comparison With In Vivo Models. *J Gerontol A Biol Sci Med Sci*.
76. Loeffler DA, DeMaggio AJ, Juneau PL, et al. (1994) Effects of enhanced striatal dopamine turnover in vivo on glutathione oxidation. *Clin Neuropharmacol* 17: 370-379.
77. Luo XG, Ding JQ, Chen SD (2010) Microglia in the aging brain: relevance to neurodegeneration. *Mol Neurodegener* 5: 12.
78. Damani MR, Zhao L, Fontainhas AM, et al. (2011) Age-related alterations in the dynamic behavior of microglia. *Aging Cell* 10: 263-276.
79. Sierra A, Gottfried-Blackmore AC, McEwen BS, et al. (2007) Microglia derived from aging mice exhibit an altered inflammatory profile. *Glia* 55: 412-424.
80. Streit WJ, Sammons NW, Kuhns AJ, et al. (2004) Dystrophic microglia in the aging human brain. *Glia* 45: 208-212.
81. Wasserman JK, Yang H, Schlichter LC (2008) Glial responses, neuron death and lesion resolution after intracerebral hemorrhage in young vs. aged rats. *Eur J Neurosci* 28: 1316-1328.
82. Streit WJ, Graeber MB (1993) Heterogeneity of microglial and perivascular cell populations: insights gained from the facial nucleus paradigm. *Glia* 7: 68-74.
83. Streit WJ, Braak H, Xue QS, et al. (2009) Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in Alzheimer's disease. *Acta Neuropathol* 118: 475-485.
84. Maezawa I, Zimin PI, Wulff H, et al. (2011) Amyloid-beta protein oligomer at low nanomolar concentrations activates microglia and induces microglial neurotoxicity. *J Biol Chem* 286: 3693-3706.
85. Williamson LL, Sholar PW, Mistry RS, et al. (2011) Microglia and memory: modulation by early-life infection. *J Neurosci* 31: 15511-15521.
86. Varnum MM, Ikezu T (2012) The classification of microglial activation phenotypes on neurodegeneration and regeneration in Alzheimer's disease brain. *Arch Immunol Ther Exp (Warsz)* 60: 251-266.
87. Solito E, Sastre M (2012) Microglia function in Alzheimer's disease. *Front Pharmacol* 3: 14.
88. Wojtera M, Sobow T, Kloszewska I, et al. (2012) Expression of immunohistochemical markers on microglia in Creutzfeldt-Jakob disease and Alzheimer's disease: morphometric study and review of the literature. *Folia Neuropathol* 50: 74-84.
89. Graeber MB, Streit WJ (2010) Microglia: biology and pathology. *Acta Neuropathol* 119: 89-105.

90. Njie EG, Boelen E, Stassen FR, et al. (2012) Ex vivo cultures of microglia from young and aged rodent brain reveal age-related changes in microglial function. *Neurobiol Aging* 33: 195 e191-112.
91. Weggen S, Eriksen JL, Das P, et al. (2001) A subset of NSAIDs lower amyloidogenic Abeta42 independently of cyclooxygenase activity. *Nature* 414: 212-216.
92. Martin BK, Szekely C, Brandt J, et al. (2008) Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): results of a randomized, controlled trial of naproxen and celecoxib. *Arch Neurol* 65: 896-905.
93. Floden AM, Combs CK (2011) Microglia demonstrate age-dependent interaction with amyloid-beta fibrils. *J Alzheimers Dis* 25: 279-293.
94. Caldeira C, Oliveira AF, Cunha C, et al. (2014) Microglia change from a reactive to an age-like phenotype with the time in culture. *Front Cell Neurosci* 8: 152.
95. Akbar AN, Henson SM (2011) Are senescence and exhaustion intertwined or unrelated processes that compromise immunity? *Nat Rev Immunol* 11: 289-295.
96. Augert A, D. B (2013) Immunosenescence and Senescence Immunosurveillance: One of the Possible Links Explaining the Cancer Incidence in Ageing Population. In: (Ed.) DWZ, editor. *Senescence and Senescence-Related Disorders*: InTech. pp. 87-111.
97. Currais A (2015) Ageing and inflammation - A central role for mitochondria in brain health and disease. *Ageing Res Rev* 21: 30-42.
98. Baylis D, Bartlett DB, Patel HP, et al. (2013) Understanding how we age: insights into inflammaging. *Longev Healthspan* 2: 8.
99. Olivieri F, Rippo MR, Monsurro V, et al. (2013) MicroRNAs linking inflamm-aging, cellular senescence and cancer. *Ageing Res Rev* 12: 1056-1068.
100. Sheridan GK, Murphy KJ (2013) Neuron-glia crosstalk in health and disease: fractalkine and CX3CR1 take centre stage. *Open Biol* 3: 130181.
101. Cuanalo-Contreras K, Mukherjee A, Soto C (2013) Role of protein misfolding and proteostasis deficiency in protein misfolding diseases and aging. *Int J Cell Biol* 2013: 638083.
102. Levine B, Kroemer G (2008) Autophagy in the pathogenesis of disease. *Cell* 132: 27-42.
103. Yamashima T (2013) Reconsider Alzheimer's disease by the 'calpain-cathepsin hypothesis'--a perspective review. *Prog Neurobiol* 105: 1-23.
104. Guo JL, Lee VM (2014) Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases. *Nat Med* 20: 130-138.
105. Baixauli F, Lopez-Otin C, Mittelbrunn M (2014) Exosomes and autophagy: coordinated mechanisms for the maintenance of cellular fitness. *Front Immunol* 5: 403.
106. Lipinski MM, Zheng B, Lu T, et al. (2010) Genome-wide analysis reveals mechanisms modulating autophagy in normal brain aging and in Alzheimer's disease. *Proc Natl Acad Sci U S A* 107: 14164-14169.
107. Lehmann BD, Paine MS, Brooks AM, et al. (2008) Senescence-associated exosome release from human prostate cancer cells. *Cancer Res* 68: 7864-7871.
108. Trifunovic A, Larsson NG (2008) Mitochondrial dysfunction as a cause of ageing. *J Intern Med* 263: 167-178.
109. Cui H, Kong Y, Zhang H (2012) Oxidative stress, mitochondrial dysfunction, and aging. *J Signal Transduct* 2012: 646354.

110. Chistiakov DA, Sobenin IA, Revin VV, et al. (2014) Mitochondrial aging and age-related dysfunction of mitochondria. *Biomed Res Int* 2014: 238463.
111. Twig G, Shirihai OS (2011) The interplay between mitochondrial dynamics and mitophagy. *Antioxid Redox Signal* 14: 1939-1951.
112. Palikaras K, Tavernarakis N (2012) Mitophagy in neurodegeneration and aging. *Front Genet* 3: 297.
113. Liu L, Sakakibara K, Chen Q, et al. (2014) Receptor-mediated mitophagy in yeast and mammalian systems. *Cell Res* 24: 787-795.
114. Zhang J (2013) Autophagy and Mitophagy in Cellular Damage Control. *Redox Biol* 1: 19-23.
115. Chao JF, Leung Y, Wang MF, et al. (2012) Nutraceuticals and their preventive or potential therapeutic value in Parkinson's disease. *Nutr Rev* 70: 373-386.
116. Thakurta IG, Chattopadhyay M, Ghosh A, et al. (2012) Dietary supplementation with N-acetyl cysteine, alpha-tocopherol and alpha-lipoic acid reduces the extent of oxidative stress and proinflammatory state in aged rat brain. *Biogerontology* 13: 479-488.
117. Navarro A, Bandez MJ, Lopez-Cepero JM, et al. (2011) High doses of vitamin E improve mitochondrial dysfunction in rat hippocampus and frontal cortex upon aging. *Am J Physiol-Reg I* 300: R827-R834.
118. Ames BN (2010) Optimal micronutrients delay mitochondrial decay and age-associated diseases. *Mech Ageing Dev* 131: 473-479.
119. Bossy-Wetzell E, Barsoum MJ, Godzik A, et al. (2003) Mitochondrial fission in apoptosis, neurodegeneration and aging. *Curr Opin Cell Biol* 15: 706-716.
120. Tatsuta T, Langer T (2008) Quality control of mitochondria: protection against neurodegeneration and ageing. *EMBO J* 27: 306-314.
121. Rubinsztein DC, Marino G, Kroemer G (2011) Autophagy and aging. *Cell* 146: 682-695.
122. Salminen A, Ojala J, Kaarniranta K (2011) Apoptosis and aging: increased resistance to apoptosis enhances the aging process. *Cell Mol Life Sci* 68: 1021-1031.
123. Uittenbogaard M, Chiaramello A (2014) Mitochondrial biogenesis: a therapeutic target for neurodevelopmental disorders and neurodegenerative diseases. *Curr Pharm Des* 20: 5574-5593.
124. Onyango IG, Lu J, Rodova M, et al. (2010) Regulation of neuron mitochondrial biogenesis and relevance to brain health. *Biochim Biophys Acta* 1802: 228-234.
125. Xu R, Hu Q, Ma Q, et al. (2014) The protease Omi regulates mitochondrial biogenesis through the GSK3 beta/PGC-1 alpha pathway. *Cell Death Dis* 5: e1373.
126. Llorens-Martin M, Jurado J, Hernandez F, et al. (2014) GSK-3beta, a pivotal kinase in Alzheimer disease. *Front Mol Neurosci* 7: 46.
127. Maixner DW, Weng HR (2013) The Role of Glycogen Synthase Kinase 3 Beta in Neuroinflammation and Pain. *J Pharm Pharmacol (Los Angel)* 1: 001.
128. Zhou X, Zhou J, Li X, et al. (2011) GSK-3beta inhibitors suppressed neuroinflammation in rat cortex by activating autophagy in ischemic brain injury. *Biochem Biophys Res Commun* 411: 271-275.
129. Nakanishi H (2003) Neuronal and microglial cathepsins in aging and age-related diseases. *Ageing Res Rev* 2: 367-381.
130. Nixon RA (2000) A "protease activation cascade" in the pathogenesis of Alzheimer's disease. *Ann N Y Acad Sci* 924: 117-131.

131. Lee EB, Skovronsky DM, Abtahian F, et al. (2003) Secretion and intracellular generation of truncated A β in beta-site amyloid-beta precursor protein-cleaving enzyme expressing human neurons. *J Biol Chem* 278: 4458-4466.
132. Schechter I, Ziv E (2011) Cathepsins S, B and L with aminopeptidases display beta-secretase activity associated with the pathogenesis of Alzheimer's disease. *Biol Chem* 392: 555-569.
133. Zhang YW, Thompson R, Zhang H, et al. (2011) APP processing in Alzheimer's disease. *Mol Brain* 4: 3.
134. Munger JS, Haass C, Lemere CA, et al. (1995) Lysosomal processing of amyloid precursor protein to A β peptides: a distinct role for cathepsin S. *Biochem J* 311 (Pt 1): 299-305.
135. Turk V, Stoka V, Vasiljeva O, et al. (2012) Cysteine cathepsins: from structure, function and regulation to new frontiers. *Biochim Biophys Acta* 1824: 68-88.
136. Gan L, Ye S, Chu A, et al. (2004) Identification of cathepsin B as a mediator of neuronal death induced by A β -activated microglial cells using a functional genomics approach. *J Biol Chem* 279: 5565-5572.
137. Petanceska S, Canoll P, Devi LA (1996) Expression of rat cathepsin S in phagocytic cells. *J Biol Chem* 271: 4403-4409.
138. Guicciardi ME, Deussing J, Miyoshi H, et al. (2000) Cathepsin B contributes to TNF-alpha-mediated hepatocyte apoptosis by promoting mitochondrial release of cytochrome c. *J Clin Invest* 106: 1127-1137.
139. Stoka V, Turk B, Schendel SL, et al. (2001) Lysosomal protease pathways to apoptosis. Cleavage of bid, not pro-caspases, is the most likely route. *J Biol Chem* 276: 3149-3157.
140. Boya P, Andreau K, Poncet D, et al. (2003) Lysosomal membrane permeabilization induces cell death in a mitochondrion-dependent fashion. *J Exp Med* 197: 1323-1334.
141. Conus S, Perozzo R, Reinheckel T, et al. (2008) Caspase-8 is activated by cathepsin D initiating neutrophil apoptosis during the resolution of inflammation. *J Exp Med* 205: 685-698.
142. Blomgran R, Zheng L, Stendahl O (2007) Cathepsin-cleaved Bid promotes apoptosis in human neutrophils via oxidative stress-induced lysosomal membrane permeabilization. *J Leukoc Biol* 81: 1213-1223.
143. Lemere CA, Munger JS, Shi GP, et al. (1995) The lysosomal cysteine protease, cathepsin S, is increased in Alzheimer's disease and Down syndrome brain. An immunocytochemical study. *Am J Pathol* 146: 848-860.
144. Wendt W, Lubbert H, Stichel CC (2008) Upregulation of cathepsin S in the aging and pathological nervous system of mice. *Brain Res* 1232: 7-20.
145. Barclay J, Clark AK, Ganju P, et al. (2007) Role of the cysteine protease cathepsin S in neuropathic hyperalgesia. *Pain* 130: 225-234.
146. Clark AK, Grist J, Al-Kashi A, et al. (2012) Spinal cathepsin S and fractalkine contribute to chronic pain in the collagen-induced arthritis model. *Arthritis Rheum* 64: 2038-2047.
147. Xu J, Wang H, Ding K, et al. (2013) Inhibition of cathepsin S produces neuroprotective effects after traumatic brain injury in mice. *Mediators Inflamm* 2013: 187873.
148. Clark AK, Yip PK, Malcangio M (2009) The liberation of fractalkine in the dorsal horn requires microglial cathepsin S. *J Neurosci* 29: 6945-6954.
149. Clark AK, Malcangio M (2012) Microglial signalling mechanisms: Cathepsin S and Fractalkine. *Exp Neurol* 234: 283-292.

150. Clark AK, Yip PK, Grist J, et al. (2007) Inhibition of spinal microglial cathepsin S for the reversal of neuropathic pain. *Proc Natl Acad Sci U S A* 104: 10655-10660.
151. Cribbs DH, Berchtold NC, Perreau V, et al. (2012) Extensive innate immune gene activation accompanies brain aging, increasing vulnerability to cognitive decline and neurodegeneration: a microarray study. *J Neuroinflammation* 9: 179.
152. Cho SH, Sun B, Zhou Y, et al. (2011) CX3CR1 protein signaling modulates microglial activation and protects against plaque-independent cognitive deficits in a mouse model of Alzheimer disease. *J Biol Chem* 286: 32713-32722.
153. Rogers JT, Morganti JM, Bachstetter AD, et al. (2011) CX3CR1 deficiency leads to impairment of hippocampal cognitive function and synaptic plasticity. *J Neurosci* 31: 16241-16250.
154. Bachstetter AD, Morganti JM, Jernberg J, et al. (2011) Fractalkine and CX 3 CR1 regulate hippocampal neurogenesis in adult and aged rats. *Neurobiol Aging* 32: 2030-2044.
155. Wynne AM, Henry CJ, Huang Y, et al. (2010) Protracted downregulation of CX3CR1 on microglia of aged mice after lipopolysaccharide challenge. *Brain Behav Immun* 24: 1190-1201.
156. Kim TS, Lim HK, Lee JY, et al. (2008) Changes in the levels of plasma soluble fractalkine in patients with mild cognitive impairment and Alzheimer's disease. *Neurosci Lett* 436: 196-200.
157. Lee S, Varvel NH, Konerth ME, et al. (2010) CX3CR1 deficiency alters microglial activation and reduces beta-amyloid deposition in two Alzheimer's disease mouse models. *Am J Pathol* 177: 2549-2562.
158. Noda M, Doi Y, Liang J, et al. (2011) Fractalkine attenuates excito-neurotoxicity via microglial clearance of damaged neurons and antioxidant enzyme heme oxygenase-1 expression. *J Biol Chem* 286: 2308-2319.
159. Bolmont T, Haiss F, Eicke D, et al. (2008) Dynamics of the microglial/amyloid interaction indicate a role in plaque maintenance. *J Neurosci* 28: 4283-4292.
160. Mrak RE (2012) Microglia in Alzheimer brain: a neuropathological perspective. *Int J Alzheimers Dis* 2012: 165021.
161. Condello C, Yuan P, Schain A, et al. (2015) Microglia constitute a barrier that prevents neurotoxic protofibrillar Abeta42 hotspots around plaques. *Nat Commun* 6: 6176.
162. Liu Z, Condello C, Schain A, et al. (2010) CX3CR1 in microglia regulates brain amyloid deposition through selective protofibrillar amyloid-beta phagocytosis. *J Neurosci* 30: 17091-17101.
163. Li C, Zhao R, Gao K, et al. (2011) Astrocytes: implications for neuroinflammatory pathogenesis of Alzheimer's disease. *Curr Alzheimer Res* 8: 67-80.
164. Zhao J, O'Connor T, Vassar R (2011) The contribution of activated astrocytes to Abeta production: implications for Alzheimer's disease pathogenesis. *J Neuroinflammation* 8: 150.
165. Persengiev SP, Kondova, II, Bontrop RE (2012) The Impact of MicroRNAs on Brain Aging and Neurodegeneration. *Curr Gerontol Geriatr Res* 2012: 359369.
166. Basaiawmoit RV, Rattan SIS (2010) Cellular stress and protein misfolding during aging. In: Bross P, Gregersen N, editors. *Protein Misfolding and Cellular Stress in Disease and Aging Concepts and Protocols*. Springer, New York: Humana Press. pp. 107-117.
167. Brignull HR, Morley JF, Morimoto RI (2007) The stress of misfolded proteins: C. elegans models for neurodegenerative disease and aging. In: Csermely P, Laszlo V, editors. *Advances in Experimental Medicine and Biology - Molecular Aspects of the Stress Response: Chaperones, Membranes and Networks*. New York, U.S.A.: Landes Bioscience/Eurekah.com.

168. Hipp MS, Park SH, Hartl FU (2014) Proteostasis impairment in protein-misfolding and -aggregation diseases. *Trends Cell Biol* 24: 506-514.
169. Morimoto RI (2008) Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. *Genes Dev* 22: 1427-1438.
170. Soti C, Csermely P (2003) Aging and molecular chaperones. *Exp Gerontol* 38: 1037-1040.
171. Clos J, Westwood JT, Becker PB, et al. (1990) Molecular cloning and expression of a hexameric *Drosophila* heat shock factor subject to negative regulation. *Cell* 63: 1085-1097.
172. Cuervo AM, Wong E (2014) Chaperone-mediated autophagy: roles in disease and aging. *Cell Res* 24: 92-104.
173. Vasudevan S, Tong Y, Steitz JA (2007) Switching from repression to activation: microRNAs can up-regulate translation. *Science* 318: 1931-1934.
174. Roberts TC (2014) The MicroRNA Biology of the Mammalian Nucleus. *Mol Ther Nucleic Acids* 3: e188.
175. Jung HJ, Suh Y (2012) MicroRNA in Aging: From Discovery to Biology. *Curr Genomics* 13: 548-557.
176. Smith-Vikos T, Slack FJ (2012) MicroRNAs and their roles in aging. *J Cell Sci* 125: 7-17.
177. Inukai S, de Lencastre A, Turner M, et al. (2012) Novel microRNAs differentially expressed during aging in the mouse brain. *PLoS One* 7: e40028.
178. Zhang H, Yang H, Zhang C, et al. (2015) Investigation of microRNA expression in human serum during the aging process. *J Gerontol A Biol Sci Med Sci* 70: 102-109.
179. Lehrbach NJ, Castro C, Murfitt KJ, et al. (2012) Post-developmental microRNA expression is required for normal physiology, and regulates aging in parallel to insulin/IGF-1 signaling in *C. elegans*. *RNA* 18: 2220-2235.
180. Jiang M, Xiang Y, Wang D, et al. (2012) Dysregulated expression of miR-146a contributes to age-related dysfunction of macrophages. *Aging Cell* 11: 29-40.
181. Olivieri F, Rippo MR, Procopio AD, et al. (2013) Circulating inflamma-miRs in aging and age-related diseases. *Front Genet* 4: 121.
182. Fichtlscherer S, De Rosa S, Fox H, et al. (2010) Circulating microRNAs in patients with coronary artery disease. *Circ Res* 107: 677-684.
183. Noren Hooten N, Abdelmohsen K, Gorospe M, et al. (2010) microRNA expression patterns reveal differential expression of target genes with age. *PLoS One* 5: e10724.
184. Ponomarev ED, Veremeyko T, Weiner HL (2013) MicroRNAs are universal regulators of differentiation, activation, and polarization of microglia and macrophages in normal and diseased CNS. *Glia* 61: 91-103.
185. Saba R, Gushue S, Huzarewich RL, et al. (2012) MicroRNA 146a (miR-146a) is over-expressed during prion disease and modulates the innate immune response and the microglial activation state. *PLoS One* 7: e30832.
186. Lukiw WJ (2007) Micro-RNA speciation in fetal, adult and Alzheimer's disease hippocampus. *Neuroreport* 18: 297-300.
187. Sethi P, Lukiw WJ (2009) Micro-RNA abundance and stability in human brain: specific alterations in Alzheimer's disease temporal lobe neocortex. *Neurosci Lett* 459: 100-104.
188. Hebert SS, Horre K, Nicolai L, et al. (2008) Loss of microRNA cluster miR-29a/b-1 in sporadic Alzheimer's disease correlates with increased BACE1/beta-secretase expression. *Proc Natl Acad Sci U S A* 105: 6415-6420.

189. Guedes JR, Custodia CM, Silva RJ, et al. (2014) Early miR-155 upregulation contributes to neuroinflammation in Alzheimer's disease triple transgenic mouse model. *Hum Mol Genet* 23: 6286-6301.
190. Janelins MC, Mastrangelo MA, Oddo S, et al. (2005) Early correlation of microglial activation with enhanced tumor necrosis factor-alpha and monocyte chemoattractant protein-1 expression specifically within the entorhinal cortex of triple transgenic Alzheimer's disease mice. *J Neuroinflammation* 2: 23.
191. Provost P (2010) MicroRNAs as a molecular basis for mental retardation, Alzheimer's and prion diseases. *Brain Res* 1338: 58-66.
192. Bang C, Thum T (2012) Exosomes: new players in cell-cell communication. *Int J Biochem Cell Biol* 44: 2060-2064.
193. Thery C, Regnault A, Garin J, et al. (1999) Molecular characterization of dendritic cell-derived exosomes. Selective accumulation of the heat shock protein hsc73. *J Cell Biol* 147: 599-610.
194. Llorente A, Skotland T, Sylvanne T, et al. (2013) Molecular lipidomics of exosomes released by PC-3 prostate cancer cells. *Biochim Biophys Acta* 1831: 1302-1309.
195. Valadi H, Ekstrom K, Bossios A, et al. (2007) Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol* 9: 654-659.
196. Yoon YJ, Kim OY, Gho YS (2014) Extracellular vesicles as emerging intercellular comunicasomes. *BMB Rep* 47: 531-539.
197. Pan BT, Teng K, Wu C, et al. (1985) Electron microscopic evidence for externalization of the transferrin receptor in vesicular form in sheep reticulocytes. *J Cell Biol* 101: 942-948.
198. Lee Y, El Andaloussi S, Wood MJ (2012) Exosomes and microvesicles: extracellular vesicles for genetic information transfer and gene therapy. *Hum Mol Genet* 21: R125-134.
199. Kastelowitz N, Yin H (2014) Exosomes and microvesicles: identification and targeting by particle size and lipid chemical probes. *Chembiochem* 15: 923-928.
200. Villarroya-Beltri C, Gutierrez-Vazquez C, Sanchez-Cabo F, et al. (2013) Sumoylated hnRNPA2B1 controls the sorting of miRNAs into exosomes through binding to specific motifs. *Nat Commun* 4: 2980.
201. Cocucci E, Meldolesi J (2015) Ectosomes and exosomes: shedding the confusion between extracellular vesicles. *Trends Cell Biol* 25: 364-372.
202. Choi DS, Kim DK, Kim YK, et al. (2014) Proteomics of extracellular vesicles: Exosomes and ectosomes. *Mass Spectrom Rev* 34: 474-490.
203. Cao J (2014) The functional role of long non-coding RNAs and epigenetics. *Biol Proced Online* 16: 11.
204. Gezer U, Ozgur E, Cetinkaya M, et al. (2014) Long non-coding RNAs with low expression levels in cells are enriched in secreted exosomes. *Cell Biol Int* 38: 1076-1079.
205. Mittelbrunn M, Gutierrez-Vazquez C, Villarroya-Beltri C, et al. (2011) Unidirectional transfer of microRNA-loaded exosomes from T cells to antigen-presenting cells. *Nat Commun* 2: 282.
206. Gupta A, Pulliam L (2014) Exosomes as mediators of neuroinflammation. *J Neuroinflammation* 11: 68.
207. Potolicchio I, Carven GJ, Xu X, et al. (2005) Proteomic analysis of microglia-derived exosomes: metabolic role of the aminopeptidase CD13 in neuropeptide catabolism. *J Immunol* 175: 2237-2243.

208. Turola E, Furlan R, Bianco F, et al. (2012) Microglial microvesicle secretion and intercellular signaling. *Front Physiol* 3: 149.
209. Danzer KM, Kranich LR, Ruf WP, et al. (2012) Exosomal cell-to-cell transmission of alpha synuclein oligomers. *Mol Neurodegener* 7: 42.
210. Banigan MG, Kao PF, Kozubek JA, et al. (2013) Differential expression of exosomal microRNAs in prefrontal cortices of schizophrenia and bipolar disorder patients. *PLoS One* 8: e48814.
211. Beveridge NJ, Cairns MJ (2012) MicroRNA dysregulation in schizophrenia. *Neurobiol Dis* 46: 263-271.
212. Mellios N, Sur M (2012) The Emerging Role of microRNAs in Schizophrenia and Autism Spectrum Disorders. *Front Psychiatry* 3: 39.
213. Mundalil Vasu M, Anitha A, Thanseem I, et al. (2014) Serum microRNA profiles in children with autism. *Mol Autism* 5: 40.
214. Dujardin S, Begard S, Caillierez R, et al. (2014) Ectosomes: a new mechanism for non-exosomal secretion of tau protein. *PLoS One* 9: e100760.
215. Rajendran L, Honsho M, Zahn TR, et al. (2006) Alzheimer's disease beta-amyloid peptides are released in association with exosomes. *Proc Natl Acad Sci U S A* 103: 11172-11177.
216. Yuyama K, Sun H, Mitsutake S, et al. (2012) Sphingolipid-modulated exosome secretion promotes clearance of amyloid-beta by microglia. *J Biol Chem* 287: 10977-10989.
217. Yuyama K, Sun H, Usuki S, et al. (2015) A potential function for neuronal exosomes: sequestering intracerebral amyloid-beta peptide. *FEBS Lett* 589: 84-88.
218. Joshi P, Turola E, Ruiz A, et al. (2014) Microglia convert aggregated amyloid-beta into neurotoxic forms through the shedding of microvesicles. *Cell Death Differ* 21: 582-593.
219. Joshi P, Benussi L, Furlan R, et al. (2015) Extracellular Vesicles in Alzheimer's Disease: Friends or Foes? Focus on A β -Vesicle Interaction. *Int J Mol Sci* 16: 4800-4813.
220. Bahrini I, Song JH, Diez D, et al. (2015) Neuronal exosomes facilitate synaptic pruning by up-regulating complement factors in microglia. *Sci Rep* 5: 7989.
221. Fitzner D, Schnaars M, van Rossum D, et al. (2011) Selective transfer of exosomes from oligodendrocytes to microglia by macropinocytosis. *J Cell Sci* 124: 447-458.
222. Basso M, Pozzi S, Tortarolo M, et al. (2013) Mutant copper-zinc superoxide dismutase (SOD1) induces protein secretion pathway alterations and exosome release in astrocytes: implications for disease spreading and motor neuron pathology in amyotrophic lateral sclerosis. *J Biol Chem* 288: 15699-15711.
223. Grad LI, Yerbury JJ, Turner BJ, et al. (2014) Intercellular propagated misfolding of wild-type Cu/Zn superoxide dismutase occurs via exosome-dependent and -independent mechanisms. *Proc Natl Acad Sci U S A* 111: 3620-3625.
224. Forsberg K, Jonsson PA, Andersen PM, et al. (2010) Novel antibodies reveal inclusions containing non-native SOD1 in sporadic ALS patients. *PLoS One* 5: e11552.
225. Xu D, Tahara H (2013) The role of exosomes and microRNAs in senescence and aging. *Adv Drug Deliv Rev* 65: 368-375.
226. Mitsuhashi M, Taub DD, Kapogiannis D, et al. (2013) Aging enhances release of exosomal cytokine mRNAs by A β 1-42-stimulated macrophages. *FASEB J* 27: 5141-5150.
227. Kawikova I, Askenase PW (2014) Diagnostic and therapeutic potentials of exosomes in CNS diseases. *Brain Res* [in press].

228. Properzi F, Logozzi M, Fais S (2013) Exosomes: the future of biomarkers in medicine. *Biomark Med* 7: 769-778.
229. Fiandaca MS, Kapogiannis D, Mapstone M, et al. (2014) Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: A case-control study. *Alzheimers Dement* [in press].
230. Cheng L, Doecke JD, Sharples RA, et al. (2014) Prognostic serum miRNA biomarkers associated with Alzheimer's disease shows concordance with neuropsychological and neuroimaging assessment. *Mol Psychiatry* [in press].
231. Liu CG, Song J, Zhang YQ, et al. (2014) MicroRNA-193b is a regulator of amyloid precursor protein in the blood and cerebrospinal fluid derived exosomal microRNA-193b is a biomarker of Alzheimer's disease. *Mol Med Rep* 10: 2395-2400.
232. Predecki M, Dorszewska J (2014) The Role of MicroRNA in the Pathogenesis and Diagnosis of Neurodegenerative Diseases. *Austin Alzheimers J Parkinsons Dis* 1: 10.
233. Lin J, Li J, Huang B, et al. (2015) Exosomes: novel biomarkers for clinical diagnosis. *ScientificWorldJournal* 2015: 657086.
234. Cheng L, Sharples RA, Scicluna BJ, et al. (2014) Exosomes provide a protective and enriched source of miRNA for biomarker profiling compared to intracellular and cell-free blood. *J Extracell Vesicles* 3.
235. Kalani A, Tyagi A, Tyagi N (2014) Exosomes: mediators of neurodegeneration, neuroprotection and therapeutics. *Mol Neurobiol* 49: 590-600.
236. Lai RC, Yeo RW, Tan KH, et al. (2013) Exosomes for drug delivery - a novel application for the mesenchymal stem cell. *Biotechnol Adv* 31: 543-551.
237. Limke TL, Rao MS (2003) Neural stem cell therapy in the aging brain: pitfalls and possibilities. *J Hematother Stem Cell Res* 12: 615-623.
238. Wong WT (2013) Microglial aging in the healthy CNS: phenotypes, drivers, and rejuvenation. *Front Cell Neurosci* 7: 22.
239. Lapasset L, Milhavet O, Prieur A, et al. (2011) Rejuvenating senescent and centenarian human cells by reprogramming through the pluripotent state. *Genes Dev* 25: 2248-2253.
240. Manukyan M, Singh PB (2014) Epigenome rejuvenation: HP1beta mobility as a measure of pluripotent and senescent chromatin ground states. *Sci Rep* 4: 4789.
241. Lunn JS, Sakowski SA, Hur J, et al. (2011) Stem cell technology for neurodegenerative diseases. *Ann Neurol* 70: 353-361.
242. Mattson MP, Chan SL, Duan W (2002) Modification of brain aging and neurodegenerative disorders by genes, diet, and behavior. *Physiol Rev* 82: 637-672.
243. Dantuma E, Merchant S, Sugaya K (2010) Stem cells for the treatment of neurodegenerative diseases. *Stem Cell Res Ther* 1: 37.
244. Thomsen GM, Gowing G, Svendsen S, et al. (2014) The past, present and future of stem cell clinical trials for ALS. *Exp Neurol* 262 Pt B: 127-137.
245. Sikora E (2013) Rejuvenation of senescent cells-the road to postponing human aging and age-related disease? *Exp Gerontol* 48: 661-666.
246. Miller RA, Harrison DE, Astle CM, et al. (2014) Rapamycin-mediated lifespan increase in mice is dose and sex dependent and metabolically distinct from dietary restriction. *Aging Cell* 13: 468-477.

247. Ehninger D, Neff F, Xie K (2014) Longevity, aging and rapamycin. *Cell Mol Life Sci* 71: 4325-4346.
248. Saemann MD, Haidinger M, Hecking M, et al. (2009) The multifunctional role of mTOR in innate immunity: implications for transplant immunity. *Am J Transplant* 9: 2655-2661.
249. Soliman GA (2013) The role of mechanistic target of rapamycin (mTOR) complexes signaling in the immune responses. *Nutrients* 5: 2231-2257.
250. Olivieri F, Mazzanti I, Abbatecola AM, et al. (2012) Telomere/Telomerase system: a new target of statins pleiotropic effect? *Curr Vasc Pharmacol* 10: 216-224.
251. Balu M, Sangeetha P, Haripriya D, et al. (2005) Rejuvenation of antioxidant system in central nervous system of aged rats by grape seed extract. *Neurosci Lett* 383: 295-300.
252. Acosta S, Jernberg J, Sanberg CD, et al. (2010) NT-020, a natural therapeutic approach to optimize spatial memory performance and increase neural progenitor cell proliferation and decrease inflammation in the aged rat. *Rejuvenation Res* 13: 581-588.
253. Prokopov AF (2010) A case of recovery from dementia following rejuvenative treatment. *Rejuvenation Res* 13: 217-219.
254. Pitozzi V, Jacomelli M, Catelan D, et al. (2012) Long-term dietary extra-virgin olive oil rich in polyphenols reverses age-related dysfunctions in motor coordination and contextual memory in mice: role of oxidative stress. *Rejuvenation Res* 15: 601-612.
255. Gillette-Guyonnet S, Secher M, Vellas B (2013) Nutrition and neurodegeneration: epidemiological evidence and challenges for future research. *Br J Clin Pharmacol* 75: 738-755.
256. Kidd PM (2005) Neurodegeneration from mitochondrial insufficiency: nutrients, stem cells, growth factors, and prospects for brain rebuilding using integrative management. *Altern Med Rev* 10: 268-293.
257. Maruszak A, Zekanowski C (2011) Mitochondrial dysfunction and Alzheimer's disease. *Prog Neuropsychopharmacol Biol Psychiatry* 35: 320-330.
258. Safdar A, Bourgeois JM, Ogborn DI, et al. (2011) Endurance exercise rescues progeroid aging and induces systemic mitochondrial rejuvenation in mtDNA mutator mice. *P Natl Acad Sci U S A* 108: 4135-4140.
259. Garcia-Matas S, Paul RK, Molina-Martinez P, et al. (2015) In vitro caloric restriction induces protective genes and functional rejuvenation in senescent SAMP8 astrocytes. *Aging Cell* 14: 334-344.
260. Suhr ST, Chang EA, Tjong J, et al. (2010) Mitochondrial rejuvenation after induced pluripotency. *PLoS One* 5: e14095.
261. Masotti A, Celluzzi A, Petrini S, et al. (2014) Aged iPSCs display an uncommon mitochondrial appearance and fail to undergo in vitro neurogenesis. *Aging (Albany NY)* 6: 1094-1108.
262. Johansson JU, Woodling NS, Wang Q, et al. (2015) Prostaglandin signaling suppresses beneficial microglial function in Alzheimer's disease models. *J Clin Invest* 125: 350-364.
263. Simard AR, Rivest S (2004) Bone marrow stem cells have the ability to populate the entire central nervous system into fully differentiated parenchymal microglia. *Faseb J* 18: 998-1000.
264. Jung S, Schwartz M (2012) Non-identical twins - microglia and monocyte-derived macrophages in acute injury and autoimmune inflammation. *Front Immunol* 3: 89.
265. London A, Cohen M, Schwartz M (2013) Microglia and monocyte-derived macrophages: functionally distinct populations that act in concert in CNS plasticity and repair. *Front Cell Neurosci* 7: 34.

266. Danielyan L, Beer-Hammer S, Stolzing A, et al. (2014) Intranasal delivery of bone marrow-derived mesenchymal stem cells, macrophages, and microglia to the brain in mouse models of Alzheimer's and Parkinson's disease. *Cell Transplant* 23 Suppl 1: S123-139.
267. Le Blon D, Hoornaert C, Daans J, et al. (2014) Distinct spatial distribution of microglia and macrophages following mesenchymal stem cell implantation in mouse brain. *Immunol Cell Biol* 92: 650-658.
268. Brites D, Vaz AR (2014) Microglia Centered Pathogenesis in ALS: Insights in Cell Interconnectivity. *Front Cell Neurosci* 8: 117.
269. Feldman EL, Boulis NM, Hur J, et al. (2014) Intraspinal neural stem cell transplantation in amyotrophic lateral sclerosis: phase 1 trial outcomes. *Ann Neurol* 75: 363-373.
270. Mazzini L, Gelati M, Profico DC, et al. (2015) Human neural stem cell transplantation in ALS: initial results from a phase I trial. *J Transl Med* 13: 371.
271. Diaz-Amarilla P, Olivera-Bravo S, Trias E, et al. (2011) Phenotypically aberrant astrocytes that promote motoneuron damage in a model of inherited amyotrophic lateral sclerosis. *Proc Natl Acad Sci U S A* 108: 18126-18131.
272. Meyer K, Ferraiuolo L, Miranda CJ, et al. (2014) Direct conversion of patient fibroblasts demonstrates non-cell autonomous toxicity of astrocytes to motor neurons in familial and sporadic ALS. *Proc Natl Acad Sci U S A* 111: 829-832.
273. Kondo T, Funayama M, Tsukita K, et al. (2014) Focal transplantation of human iPSC-derived glial-rich neural progenitors improves lifespan of ALS mice. *Stem Cell Reports* 3: 242-249.
274. Sareen D, Gowing G, Sahabian A, et al. (2014) Human induced pluripotent stem cells are a novel source of neural progenitor cells (iNPCs) that migrate and integrate in the rodent spinal cord. *J Comp Neurol* 522: 2707-2728.
275. Drago D, Cossetti C, Iraci N, et al. (2013) The stem cell secretome and its role in brain repair. *Biochimie* 95: 2271-2285.
276. Haas C (2012) Strategies, development, and pitfalls of therapeutic options for Alzheimer's disease. *J Alzheimers Dis* 28: 241-281.
277. Li XY, Bao XJ, Wang RZ (2015) Potential of neural stem cell-based therapies for Alzheimer's disease. *J Neurosci Res* [in press].
278. Zhang W, Wang PJ, Sha HY, et al. (2014) Neural Stem Cell Transplants Improve Cognitive Function Without Altering Amyloid Pathology in an APP/PS1 Double Transgenic Model of Alzheimer's Disease. *Mol Neurobiol* 50: 423-437.
279. Yan Y, Ma T, Gong K, et al. (2014) Adipose-derived mesenchymal stem cell transplantation promotes adult neurogenesis in the brains of Alzheimer's disease mice. *Neural Regen Res* 9: 798-805.
280. Services USDoHaH, National Plan to Address Alzheimer's Disease: 2013 Update. 2013. Available from: <http://aspe.hhs.gov/daltcp/napa/NatlPlan2013.pdf>
281. Prince M, Bryce R, Albanese E, et al. (2013) The global prevalence of dementia: a systematic review and metaanalysis. *Alzheimers Dement* 9: 63-75 e62.
282. Yang Z, Lin PJ, Levey A (2013) Monetary costs of dementia in the United States. *N Engl J Med* 369: 489.
283. Hurd MD, Martorell P, Langa KM (2013) Monetary costs of dementia in the United States. *N Engl J Med* 369: 489-490.

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284. Qiu C, Kivipelto M, von Strauss E (2009) Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. *Dialogues Clin Neurosci* 11: 111-128.
285. Ruzankina Y, Brown EJ (2007) Relationships between stem cell exhaustion, tumour suppression and ageing. *Brit J Cancer* 97: 1189-1193.
286. Conover JC, Shook BA (2011) Aging of the subventricular zone neural stem cell niche. *Aging Dis* 2: 49-63.
287. Artegiani B, Calegari F (2012) Age-related cognitive decline: can neural stem cells help us? *Aging (Albany NY)* 4: 176-186.
288. van Wijngaarden P, Franklin RJ (2013) Ageing stem and progenitor cells: implications for rejuvenation of the central nervous system. *Development* 140: 2562-2575.
289. Howlett SE, Rockwood K (2014) Ageing: develop models of frailty. *Nature* 512: 253-253.
290. Bowling A, Iliffe S (2006) Which model of successful ageing should be used? Baseline findings from a British longitudinal survey of ageing. *Age Ageing* 35: 607-614.
291. Bowling A, Dieppe P (2005) What is successful ageing and who should define it? *Brit Med J* 331: 1548-1551.
292. Vanhooren V, Libert C (2013) The mouse as a model organism in aging research: Usefulness, pitfalls and possibilities. *Ageing Res Rev* 12: 8-21.



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