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Research article

Global model for in-field monitoring of sugar content and color of melon pulp with comparative regression approach

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Abstract: The development of the global model is an important part of research involving the quality prediction of agricultural commodities using visible/near-infrared (Vis/NIR) spectroscopy due to its efficiency and effectiveness. The Vis/NIR was used in this study to develop a global model and to evaluate the sugar content and pulp color, which are the main determinants of ripeness and quality of melons. Furthermore, it also provides a comparison between linear and nonlinear regression using partial least squares regression (PLSR) and support vector machine regression (SVMR), respectively. The model accuracy was determined by ratio of performance to deviation (RPD). The results showed that there were good model accuracy values in some parameters, such as SSC (2.14), glucose (1.59), sucrose (2.31), a* (2.97), and b* (2.49), while the fructose (1.35) and L* (1.06) modeling showed poor prediction accuracy. The best model for SSC was developed using PLSR, while that of fructose, glucose, sucrose, L*, a*, and b* were obtained from SVMR. Therefore, Vis/NIR spectroscopy can be used as an alternative method to monitor sugar content and pulp color of a melon, but with some limitations, such as the low accuracy in predicting certain variables, such as the L* and fructose.

Keywords: contactless; machine learning; multivariate analysis; on-tree; pulp color

1. Introduction

Melon is one of the most popular fruits in the world, which is widely cultivated in tropical and

subtropical countries. Furthermore, it is a non-climacteric fruit since its ripening process only occurs while the fruit is still on the tree. Melon does not ripen after harvesting unlike what is seen in other fruits, such as bananas or papayas. Subsequently, ripeness has been found to be the best measure of the quality of most fruit commodities in comparison to other external properties, such as the peel shape or color, which cannot ascertain the sugar content based on the soluble solids content (SSC), fructose, glucose, and sucrose. Traditional methods for determining the SSC, fructose, glucose, and sucrose in melons are destructive, time-consuming, and unsuitable for large-scale data acquisition or field use to assess the internal quality of the fruit on a tree in real-time.

The potential development of non-destructive methods could enable the rapid and repeated analysis of large quantities on fruit samples for a variety of purposes. Various studies have been reported regarding the use of this non-destructive technology to predict water content and SSC of various commodities in the *Cucurbitaceae* family [1–3]. Additionally, this technology could be potentially used for concurrent collection of data based on different quality indices to allow the development of postharvest protocol and to separate the fruits based on internal quality [4–7].

According to the various types of technology developed in the last few decades, visible/nearinfrared (Vis/NIR) spectroscopy is a promising non-destructive method for determining quality standards in agricultural products [8]. The main focus of Vis/NIR development is the prediction and monitoring of various internal quality traits in fruits while on the tree or after harvesting [9]. Furthermore, this technology was selected to determine the sugar content in melons by the presence of important wavelengths in the visible/near-infrared region since it is safe, fast, and non-destructive. The implementation of Vis/NIR in the field allows farmers to determine the quality of fruits in trees, which eases the decision making before harvest time. Also, this procedure can be used to sort large samples in order to improve efficiency and make decisions in real-time on each distribution chain [10].

Several studies on the development of the Vis/NIR model for different varieties of single fruit species has shown that this method increases the robustness and accuracy of the developed calibration model [11,12]. Additionally, it is more preferred to use this global model in real-time since its development involves the determination of the quality attributes of melons using various varieties. The development of a single Vis/NIR model is intriguing due to the various melon varieties in Indonesia. Therefore, this study aims to (1) develop a global Vis/NIR model for predicting the sugar content and pulp color in 3 melon varieties; and (2) to apply two different chemometric approaches, which includes the partial least squares regression (PLSR) and the support vector machine regression (SVMR) as a linear regression and non-linear regression method, respectively, in order to obtain an accurate Vis/NIR model. There has been no comparative study of the two methods for monitoring the quality of melon pulps with thick pulp, even though these two methods were used to predict the quality attributes of agricultural products.

2. Materials and methods

2.1. Melon samples

The sample used in this study consisted of three melon varieties in order to produce a reliable and accurate global regression model. A total of 135 melons were used, with 45 samples from each variety, such as *Linius*, *Ignazio*, and *Dalmatian* melon varieties, which were harvested from hydroponic gardens in Garut, Indonesia. A total of two-thirds of the total sample was used for calibration model

development (90 samples), while the remaining third was used for the prediction set (45 samples) to test the previously developed calibration model. The 90 samples of the calibration set were selected from 30 samples of each variety, whereas the prediction set picked an amount of 15 samples per variety.

2.2. Vis/NIR assays

The handheld NirVana AG410 (Integrated Spectronics Pty, Ltd, Australia) was used to acquire Vis-NIR spectra with a wavelength range of 381–1065 nm and spectral sampling intervals of 3 nm. The absorbance spectra of 135 whole melon samples were acquired during the acquisition using 6 data points for each. The melon sample spectrum was calculated by averaging the values of six data collection points on the same fruit. Each fruit was numbered and labelled at the six spectral collection points immediately following the acquisition of the spectra for each variety. A Vis/NIR spectrometer was used to scan the fruit parts, which were then used to collect reference data (sugar content and color). To minimize the effect of temperature variations on the performance of the resulting model, all spectra acquisition and reference analysis were performed on the same day at a temperature of 27 °C (room temperature).

2.3. Reference assays of sugar content and fruit pulp color

The pulp color reference data was measured after the sample has been destructed. L^{*}, a^{*}, and b^{*} values were used to describe the pulp color and were measured with a CM-600d spectrophotometer (Konica Minolta, INC, Japan). L^{*} denotes the brightness value, a^{*} denotes the green–red color gradation, and b^{*} denotes the blue–yellow color gradation. At a later stage, the mean value of the six measurements in each sample was used as reference data for modelling.

The fruit was peeled, cut into small pieces, sliced and blended for 5 minutes on high speed to form a 15 mL sample of melon juice, which was then transferred to a centrifuge test tube. It was also centrifuged for 7 minutes at 4000 rpm. The centrifuge supernatant was dropped on the refractometer to obtain the sample's SSC value, which was expressed as a percentage Brix [13]. Each sample was measured three times, and the average of the three measurements was calculated. The fructose, glucose, and sucrose content were determined using high-performance liquid chromatography (Shimadzu, LC 20AT Prominence, Japan) and the procedure described by Damayanti et al. [14] and Saad et al. [15].

2.4. Chemometric models

The calibration model was created using Vis/NIR spectra and reference data on seven quality attributes: SSC, fructose, glucose, sucrose, L^{*}, a^{*}, and b^{*}. The Unscrambler X 10.4 (Camo Software AS, Oslo, Norway) was used for data analysis, including spectra preprocessing and model development. To improve the calibration model's accuracy, the original spectra were preprocessed using several methods, including standard normal variate (SNV), multiplicative scatter correction (MSC), and second derivative Savitzky-Golay (dg2). In general, preprocessing spectra help detect and correct anomalous spectra in calibration datasets, so the developed model is more accurate. SNV and MSC are popular preprocessing methods for normalizing spectra. Both methods are used to reduce the light scattering and additive effects. SNV eliminates the multiplicative effect by centering and scaling

of the original individual spectra, while MSC requires the reference spectrum that is free of scattering effects. The reference spectrum is usually employed from the average of sample data. Lastly, dg2 is effective to reduce peak overlap and linear baseline slope variations. In spectroscopic analysis, dg2 is mostly run to detect the quality attributes with low concentration. Figure 1 depicts an overview of the Vis/NIR modelling procedure. The best model for each quality attribute is selected based on the highest coefficient of determination calculation on calibration set (R²cal) and prediction set (R²pred), as well as the lowest root mean squares error of calibration set (RMSEC) and root mean squares error of prediction set (RMSEP). The results of calibration modelling involving partial least squares regression (PLSR) and support vector machine regression (SVMR), which act as chemometric algorithms in model development, yield R²cal and RMSEC. R²pred and RMSEP, on the other hand, are obtained by applying the regression equation derived from the calibration model development, which is then used to predict unknown samples. The ratio of performance to deviation (RPD) is calculated to aid in the selection of the best predictive model. RPD is calculated by dividing the standard deviation ratio in the prediction set by the RMSEP. The recommended RPD for agricultural applications is greater than 3, an RPD value between 2-3 indicates good predictive ability, 1.5-2 indicates moderate predictive ability but still needs improvement, and less than 1.5 indicates poor predictive ability [16,17].



Figure 1. Procedure for developing Vis/NIR model.

3. Results and discussion

3.1. Reference data analysis of quality attributes in three melon varieties

The results of laboratory measurements related to the quality of the three melon varieties are shown in Table 1. *Dalmatian* had the highest mean sugar content, including SSC, fructose, glucose, and sucrose, followed by *Ignazio* and *Linius* (Table 1). Additionally, the three varieties had different color values (L^* , a^* , and b^*). This is due to the orange pulp of the *Linius* melon, the white pulp of the *Ignazio* melon, and the green pulp of the *Dalmatian* melon. Because the three varieties were combined during the modelling stage, each quality attribute has a wide range, namely SSC (2.70–12.60%), fructose (1.81–7.02%), glucose (1.68–12.13%), sucrose (0.02–15.84%), L^* (29.85–74.68), a^* (–7.70–27.27), and b^* (9.57–46.42). Furthermore, this study aimed to create a global

model that can be applied to a wide range of melon varieties. Therefore, a wide range of data on quality attributes certainly aids in a better modelling process [18,19].

Figure 2 depicts the results of the principal component analysis evaluation (PCA). The reference data for *Linius* sugar content were separated from *Ignazio* and *Dalmatian* group (Figure 2(a)), which had an overlapping data distribution and indicated similarity in sugar content. However, *Dalmatian* has a higher sugar content than *Ignazio* based on the average value. The total variation in data that can be explained by three PCs is 82% (PC1), 15% (PC2), and 2% (PC3). The scatter plot for color data that includes L^{*}, a^{*}, and b^{*} in Figure 2(b) yielded values of 67%, 25%, and 8% for PC1, PC2, and PC3, respectively, with a total of 100% data variation. Unlike the sugar content scatter plot, the PCA analysis results on color data show three large groups representing each variety, with no overlapping data distribution between varieties (Figure 2(b)).

Quality attributes	Variety	Minimum	Maximum	Mean	Standard deviation
SSC (%)	Linius	2.70	8.80	4.74	1.21
	Ignazio	7.00	10.90	8.56	1.00
	Dalmatian	6.20	12.60	9.29	1.53
Fructose (%)	Linius	1.81	4.18	3.06	0.69
	Ignazio	3.05	5.95	3.95	0.63
	Dalmatian	2.99	7.02	4.49	0.84
Glucose (%)	Linius	1.68	6.80	3.79	1.20
	Ignazio	5.28	9.41	6.79	0.98
	Dalmatian	4.33	12.13	7.16	1.57
Sucrose (%)	Linius	0.02	3.30	0.84	0.77
	Ignazio	2.83	11.00	6.18	1.88
	Dalmatian	1.73	15.84	6.19	3.23
L*	Linius	29.85	68.41	57.36	8.23
	Ignazio	41.47	74.68	59.63	7.62
	Dalmatian	31.16	69.96	54.55	8.30
a [*]	Linius	12.44	27.27	19.18	3.52
	Ignazio	-1.15	8.37	1.62	2.51
	Dalmatian	-7.70	9.13	-1.84	4.15
b*	Linius	23.23	46.42	35.66	4.84
	Ignazio	9.57	22.99	14.08	3.63
	Dalmatian	13.48	41.32	23.79	5.92

Table 1. Summary of reference values in three melon varieties.



Figure 2. PCA scores plot extracted from sugar (a) and color (b) data of melon pulp.

3.2. Spectra information

The dataset contains 135 spectra from three different melon varieties. The Vis/NIR spectra in the 381–1065 nm wavelength range contain detailed information about the sample's chemical constituents. Figure 3(a) describes the example of spectrum of each variety, while Figure 3(b) displays the first two PCs of loadings plot extracted from the all acquisition of calibration set. Each variety has its own characteristics, which makes developing general quantitative models for melons difficult. Therefore, it is important to interpret the Vis/NIR spectra before the modelling stage. The original spectra revealed that the peaks at 480, 500, and 640 nm corresponded to color pigment absorptions [20,21]. Conversely, the peaks at 950 and 975 nm are the water and sugar content absorptions [22,23]. However, it is still important to optimize the original spectra to clarify the chemical information of the sample. The Vis/NIR modeling in this research was conducted to predict the quality attributes quantitatively. There is the need to eliminate the noise produced by the spectrum, as well as other irrelevant data based on the quality attributes [24,25].



Figure 3. Spectra characteristics of each melon variety (a) and loadings plot of first two PCs (b).

3.3. Chemometric calibration

Chemometric techniques in Vis/NIR modelling are broadly classified into two types: linear regression approaches and nonlinear regression approaches. PLSR is the most commonly used approach in the application of linear regression to build a calibration model on Vis/NIR data. However, PLSR has limitations in finding correlations between Vis/NIR spectral data and quality attributes in some situations. This condition occurs when the predictor and response variables have a nonlinear relationship, which can also occur when there are significant data variations in calibration modelling [26]. Therefore, this problem can be solved using a nonlinear regression approach, specifically SVMR, which has a similar sample distribution with PLSR. Additionally, the calibration set contains a broader range of data than the prediction set since it aims to develop an accurate model.

3.3.1. The PLSR approach

As described in the methods section, the SNV, MSC, and dg2 methods were used to process all of the original spectra (Figure 4). Each preprocessing spectrum was evaluated on all quality attributes (Table 2). Previous study by Wu et al. [27] reported that the use of SNV and MSC yielded highest accuracy over other spectra preprocessing for sugar content quantification in pears. Moreover, a study conducted by Xie et al. [28] regarding the prediction of glucose, fructose, and sucrose in bayberry juice concluded that second derivative preprocessing method generated the preferable result compared to the first derivative and the original spectra. Hence, this led to the application of SNV, MSC, and dg2 in our study. The combination of PLSR and dg2 was found to be the optimal model for modelling SSC, sucrose, and b^{*}. PLSR + SNV produced the most reliable model for fructose, L^{*}, and a^* prediction. On the other hand, the PLSR + original spectra combination produced the best results for the quality attribute of glucose levels. By incorporating the analyzed quality attribute data (variable Y), PLSR converts the spectral data (variable X) to a new variable (principal components), and then performs multiple linear regression on the principal component and variable Y [29]. The results of modelling with PLSR indicate that a global regression model for monitoring melon quality can be developed successfully. The following challenge is to develop a model capable of higher prediction accuracy.



Figure 4. All acquisition of original spectra (a), SNV (b), MSC (c), dg2 (d).

3.3.2. The SVMR approach

SVMR was also applied to the same data to compare its predictive accuracy with PLSR. Furthermore, SVMR can be expected to be a good candidate for this problem because of its ability to find nonlinear relationships and the good data generalizability [30,31]. The best spectra preprocessing for SSC quality attributes, fructose, glucose, sucrose, a^* , and b^* in SVMR modeling are shown by dg2 (Table 2). The dg2 method was quite effective in interpreting the low concentration of quality attributes since it could increase the spectral resolution [32]. On the other hand, SVMR + original spectra show the best calibration and prediction in L* modeling. SSC, glucose, sucrose, a^* , and b^* resulted in a reliable model to be applied in monitoring fruit quality in the field. This is indicated by the RPD values of 2.00 (SSC), 1.59 (glucose), 2.31 (sucrose), 2.97 (a^*), and 2.49 (b^*). However, SVMR showed poor predictive accuracy similar to the PLSR approach on the quality attributes of fructose and L* (Figure 5). In both models, only 1.35 (fructose) and 1.06 (L*) RPD values were obtained. This is due to the limited range of Vis/NIR wavelengths to penetrate thick-pulped melons hence the fructose and L* modeling results are less accurate (RPD < 1.5). Another cause is the variability of the physical characteristics and particle size of the sample which affects the linearity of the data.



Figure 5. Scatter plot of prediction set for each quality attributes; (a) = SSC; (b) = fructose; (c) = glucose; (d) = sucrose; (e) = L^* (lightness); (f) = a^* (green-red); (g) = b^* (blue-yellow).

Quality	Model development	Calibration		Prediction (Validation)		RPD
attributes		R ² cal	RMSEC	R ² pred	RMSEP	
SSC	PLSR + dg2	0.82	1.00	0.78	1.02	2.14
	SVMR + dg2	0.90	0.73	0.75	1.09	2.00
Fructose	PLSR + SNV	0.37	0.76	0.39	0.66	1.26
	SVMR + dg2	0.61	0.62	0.46	0.62	1.35
Glucose	PLSR + Original	0.56	1.34	0.60	1.16	1.57
	SVMR + dg2	0.63	1.24	0.61	1.15	1.59
Sucrose	PLSR + dg2	0.69	1.90	0.76	1.44	2.04
	SVMR + dg2	0.76	1.76	0.81	1.27	2.31
L^*	PLSR + SNV	0.02	8.47	0.10	7.27	1.04
	SVMR + Original	0.14	8.41	0.13	7.11	1.06
a [*]	PLSR+ SNV	0.90	3.07	0.87	3.36	2.82
	SVMR + dg2	0.93	2.50	0.88	3.19	2.97
b*	PLSR + dg2	0.78	4.74	0.80	4.20	2.26
	SVMR + dg2	0.87	3.71	0.84	3.81	2.49

Table 2. The best model for each regression approach and its preprocessing to predict quality attributes in melon.

Note: SSC: soluble solids content; PLSR: partial least squares regression; SVMR: support vector machine regression; SNV: standard normal variate; MSC: multiplicative scatter correction; R²cal: coefficient of determination in calibration set; RMSEC: root mean squares error of calibration; R²pred: coefficient of determination in prediction set; RMSEP: root mean squares error of prediction; RPD: ratio of performance to deviation.

3.3.2. Comparison of global PLSR and SVMR performances

Generally, this study aimed to promote SVMR as a viable candidate for spectral data regression. Theoretically, SVMR is advantageous for spectra regression because it is capable of performing both linear and nonlinear regression on a set of data efficiently. Basically, PLSR and SVMR can be used to predict global melon fruit quality using Vis/NIR spectroscopy [33,34]. The optimal model should exhibit the highest R²cal, R²pred, and RPD values. However, the error value of the model should be kept to a minimum (RMSEC and RMSEP). The error value of the SVMR calibration model is less than that of the PLSR. These results suggest that SVMR produces a more accurate model when dealing with variations in the data of the calibration set. This is also supported by the fact that SVMR produces a better calibration model than PLSR, as shown by the higher R²cal value on all quality attributes. On the other hand, the prediction set produced inconsistent results, which showed that the PLSR outperformed SVMR in terms of R²pred in SSC prediction. Additionally, the RMSEP of SVMR was significantly higher in SSC prediction but the reason for the increased RMSEP was still unknown. This often occurs in over-fitting models [35–37]. Conversely, SVMR produces a lower RMSEP than PLSR in other scenarios and was found to be more effective at establishing relationships between Vis/NIR spectra and complex patterns of quality attributes. However, this does not always imply that SVMR is the best method. The use of linear regression is usually preferred over nonlinear regression because of its simplicity. Additionally, certain nonlinear regression problems, such as SSC prediction in this study, can be solved using linear regression methods. SVMR may be used in place of PLSR to predict the

sugar content and pulp color of melon in a non-destructive manner, especially the three varieties of melon that are prevalent in Indonesia.

4. Conclusions

The results showed that visible near-infrared (Vis/NIR) spectroscopy can be used to predict the sugar content and pulp color of melon using a global model. Also, the resulting model showed a good accuracy based on the quality attributes of SSC, glucose, sucrose, a^* , b^* . However, the RPD values based on the quality attributes of fructose and L^{*} were 1.35 and 1.06, which were below the minimum recommended values for field application. The use of support vector machine regression (SVMR) could improve the accuracy of the model on the quality attributes of fructose, glucose, sucrose, L^* , a^* , and b^* .

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Conflict of interest

The authors declare no conflict of interest.

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